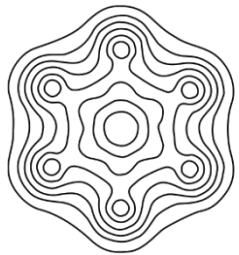
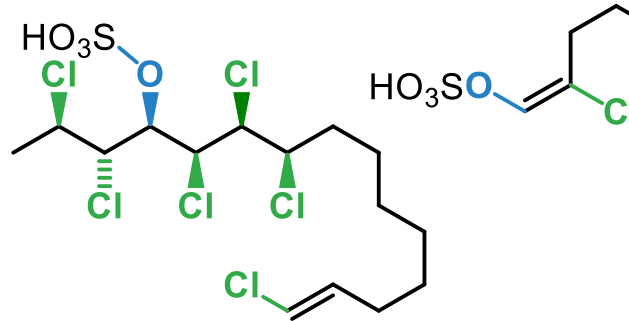
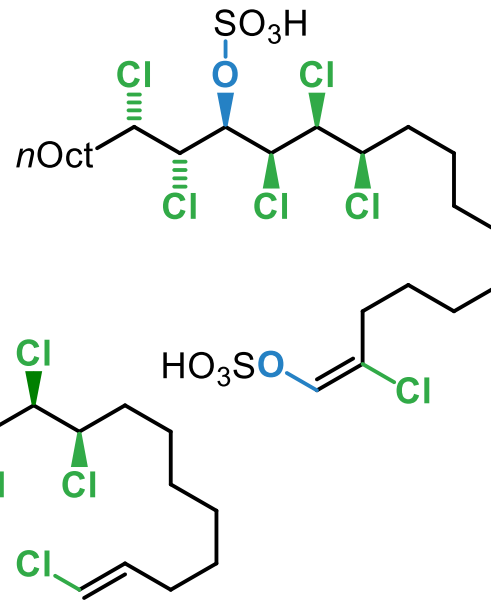
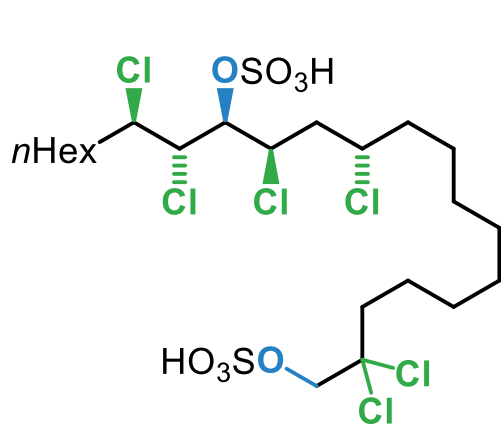


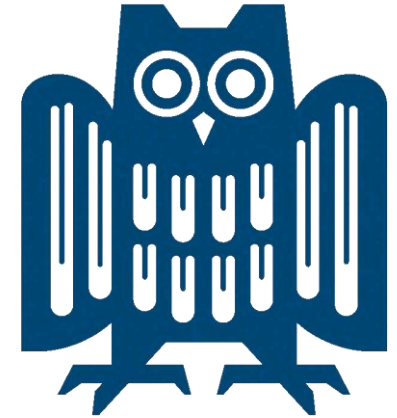
12.05.20

Wissenschaftlicher Vortrag
Im Rahmen des Habilitationsverfahrens



FCI
FONDS DER
CHEMISCHEN
INDUSTRIE

***Sulfolipide – Polyhalogenierte
toxische Naturstoff aus Algen***



DFG

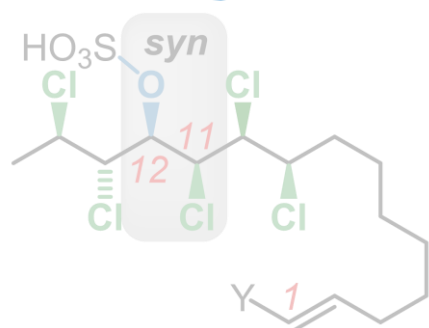
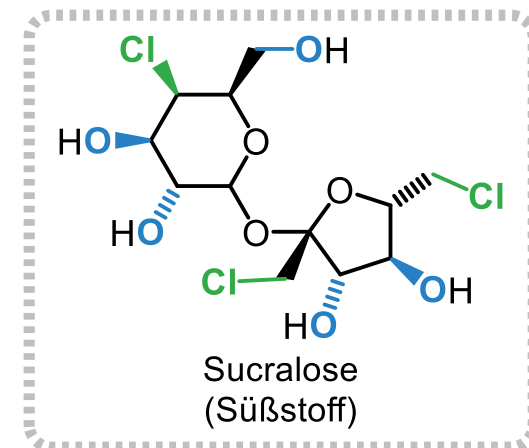
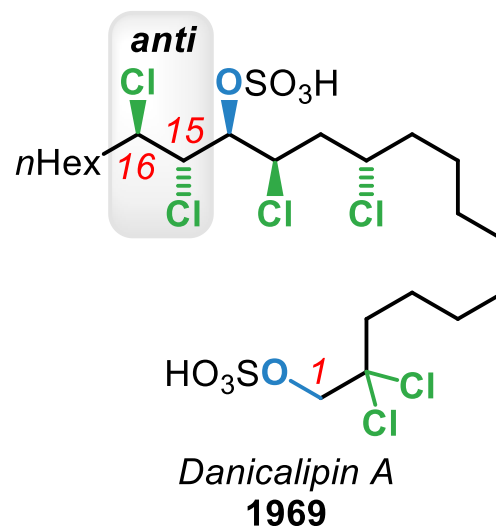
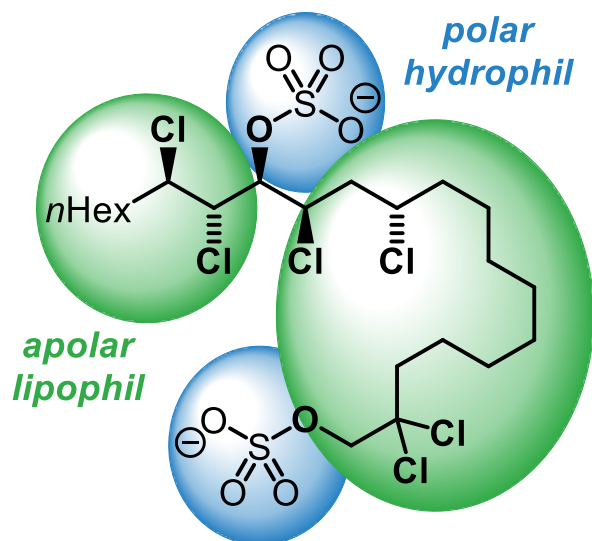
Universität des

Saarlandes

Peter Huy

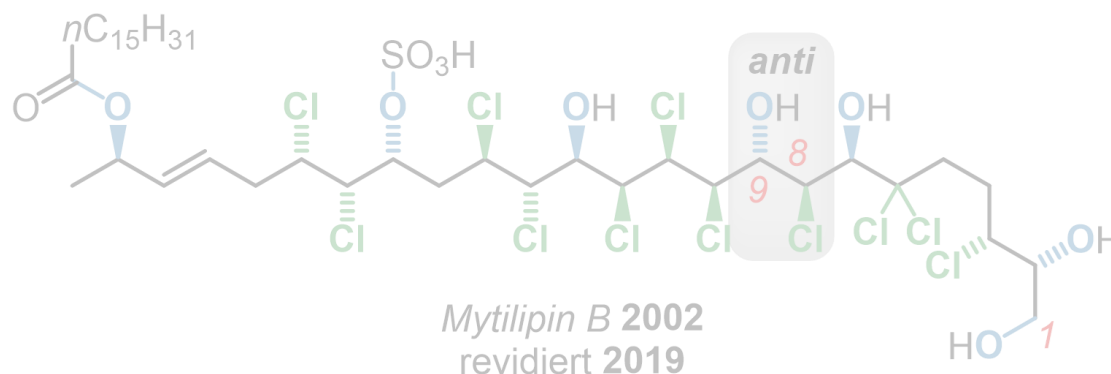
Peterhuylab.de

- Lipophil: Polychlorierte acylische Motive
- Hydrophil: Sulfonat-Gruppen → **Chlorosulfolipide**
- Isoliert aus Süßwasseralgen und Muscheln



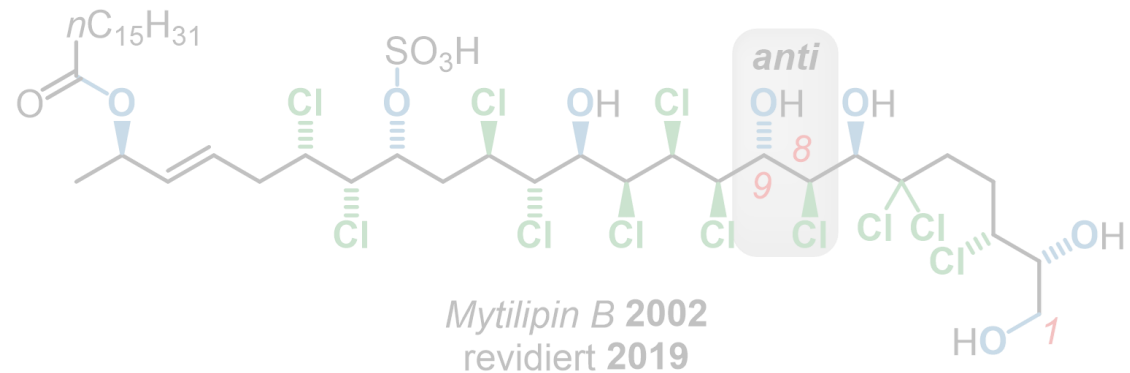
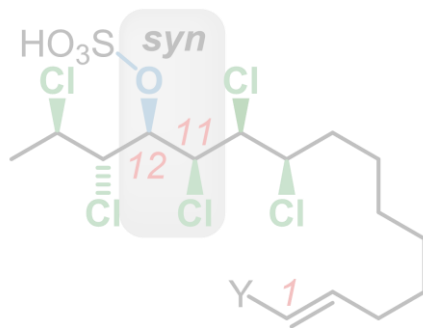
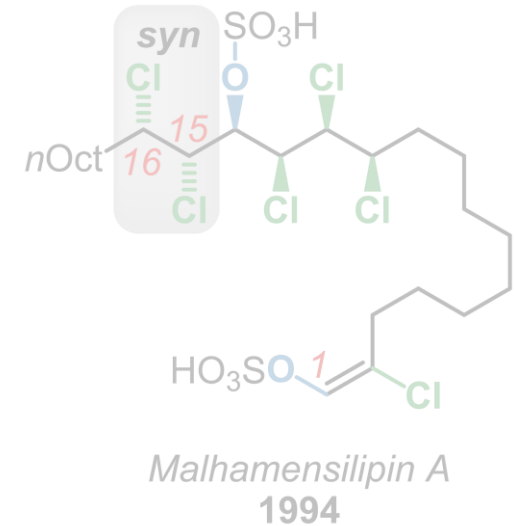
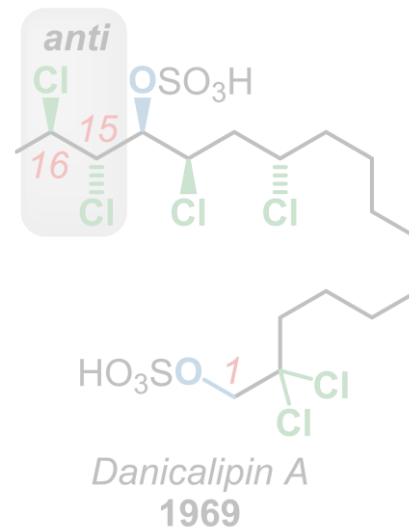
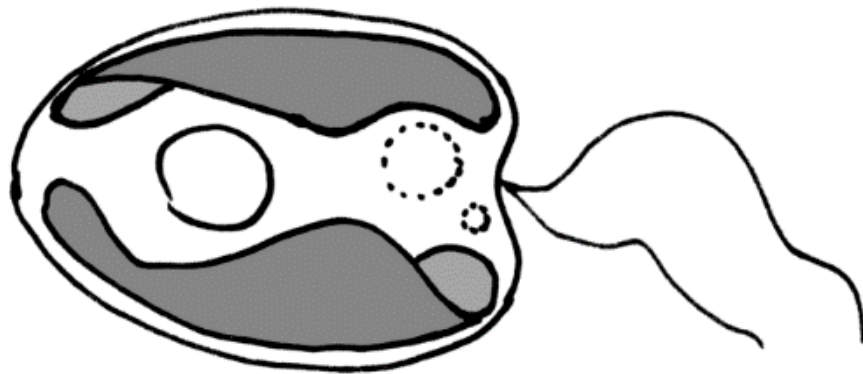
Mytilipin A 2001 (Y = Cl)

Deschloromytilipin A 2010 (Y = H)



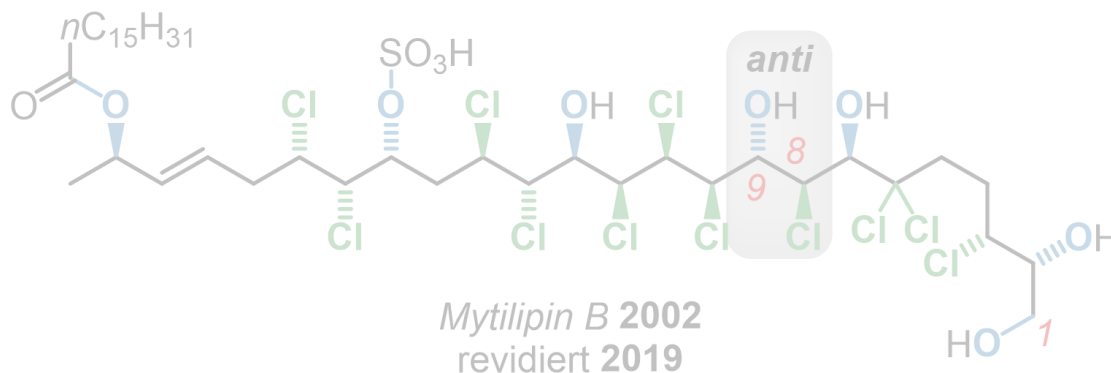
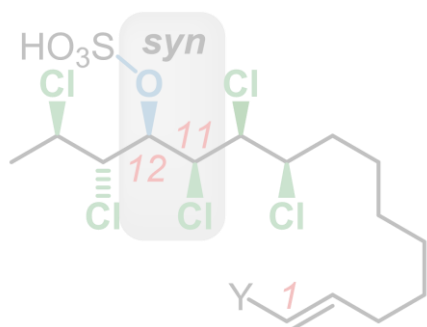
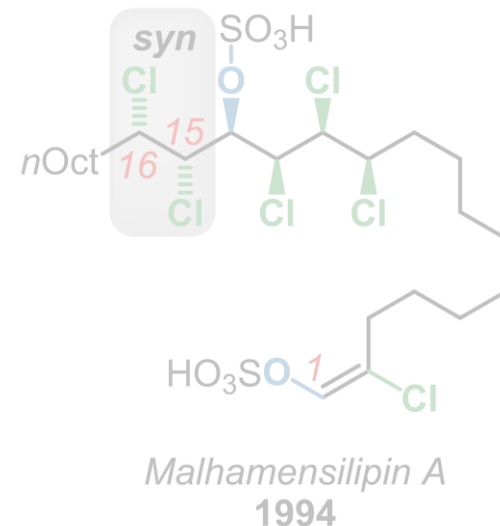
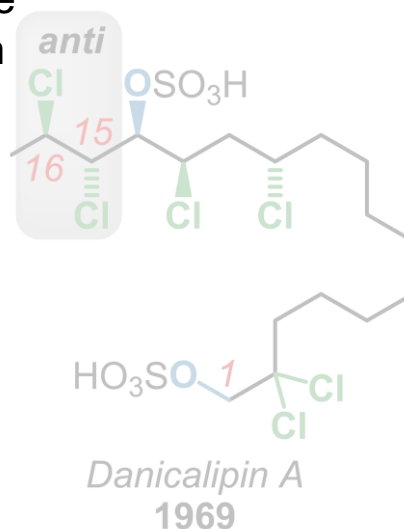
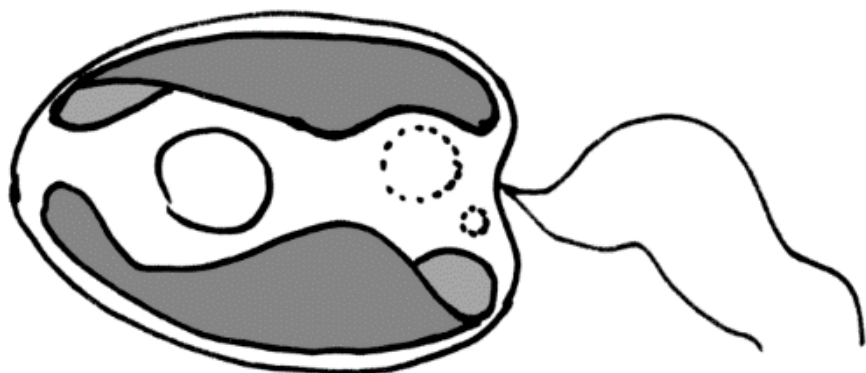
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- Hauptvorkommen goldbraunen Mikroalgen *Ochromonas danica* aus der Adriatischen See
- Hauptbestandteil der Flagellen zur Fortbewegung



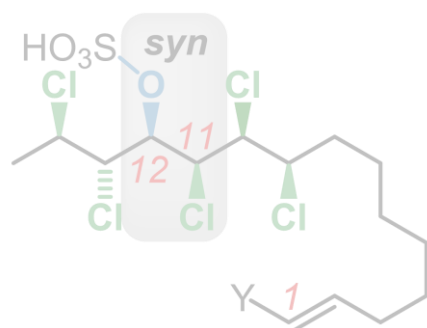
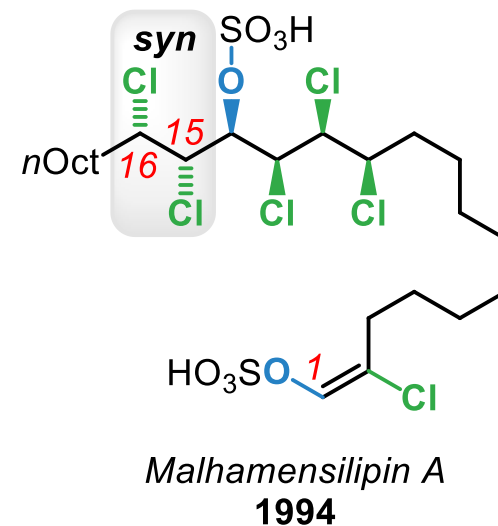
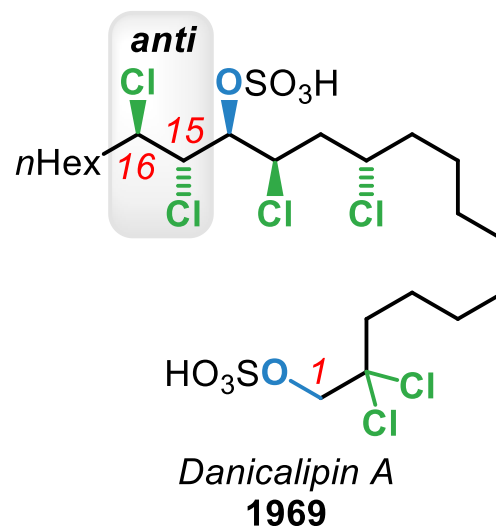
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- Hauptvorkommen goldbraunen Mikroalgen *Ochromonas danica* aus der Adriatischen See
- Hauptbestandteil der Flagellen zur Fortbewegung
- Cytotoxisch, antimikrobiell und antiproliferative
- Lebensgefährliche Meeresfrüchtevergiftungen



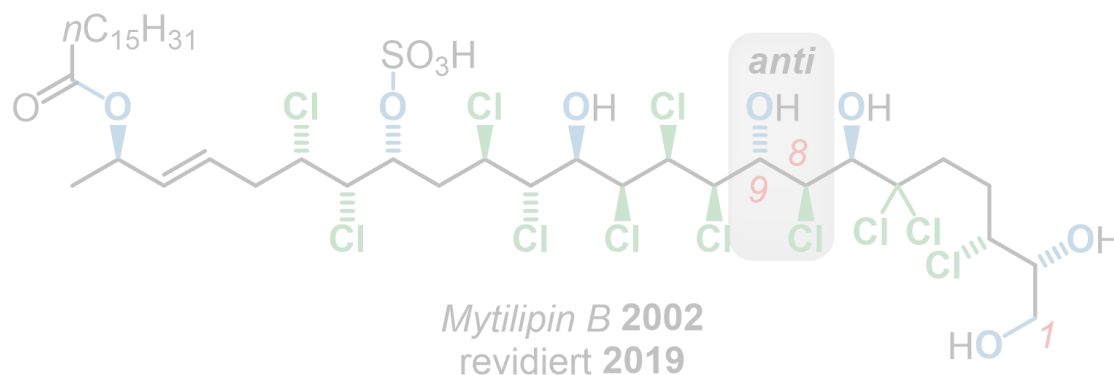
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- Hohe Stereochemische Komplexität
- *anti*- und *syn*-1,2-Dichloride



Mytilipin A 2001 (Y = Cl)

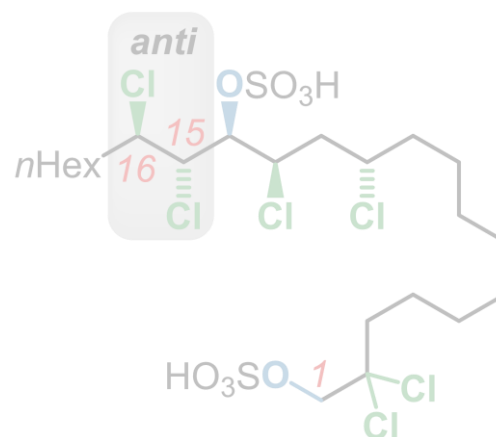
Deschloromytilipin A 2010 (Y = H)



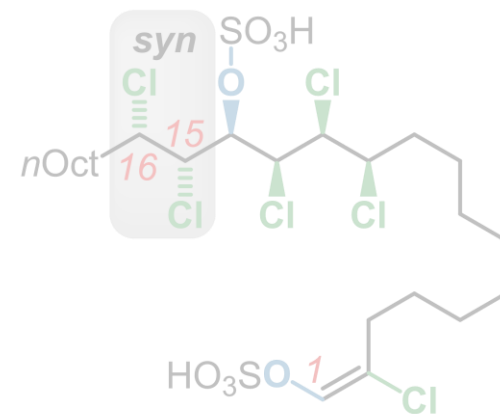
Mytilipin B 2002
revidiert 2019

Übersichtsartikel: (a) Vanderwal *et al.*, *Nat. Prod. Rep.* **2011**, 28, 15. (b) Carreira *et al.*, *Eur. J. Org. Chem.* **2012**, 1685. (c) Matsuda *et al.*, *Tetrahedron Lett.* **2014**, 55, 3003. (d) Vanderwal *et al.*, *Acc. Chem. Res.* **2014**, 47, 718. (e) P. H. Huy, P. Grewelinger, *Nachr. Chem.* **2019**, 67(9), 63.

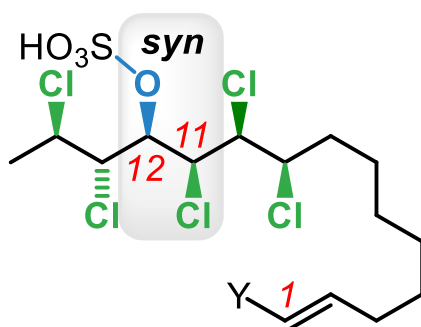
- Hohe Stereochemische Komplexität
- *anti*- und *syn*-1,2-Dichloride
- *anti*- und *syn*-Chlorhydrine
- Erste Isolation **1969**
- Erste Totalsynthese **2009**



Danicalipin A
1969

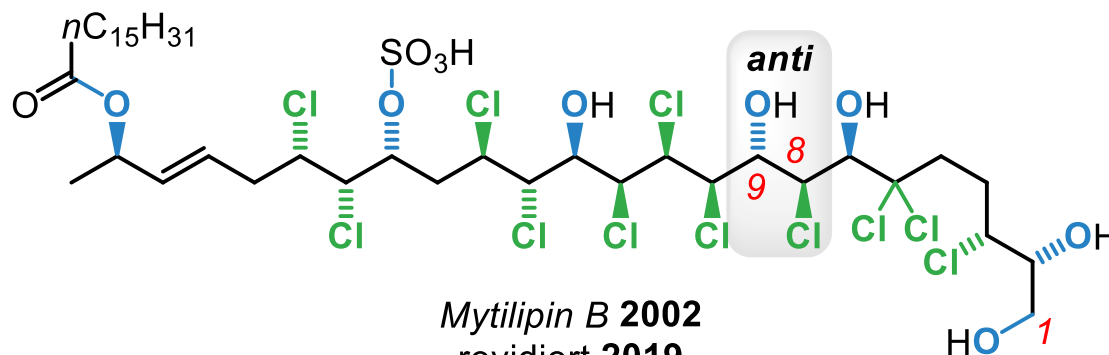


Malhamensilipin A
1994



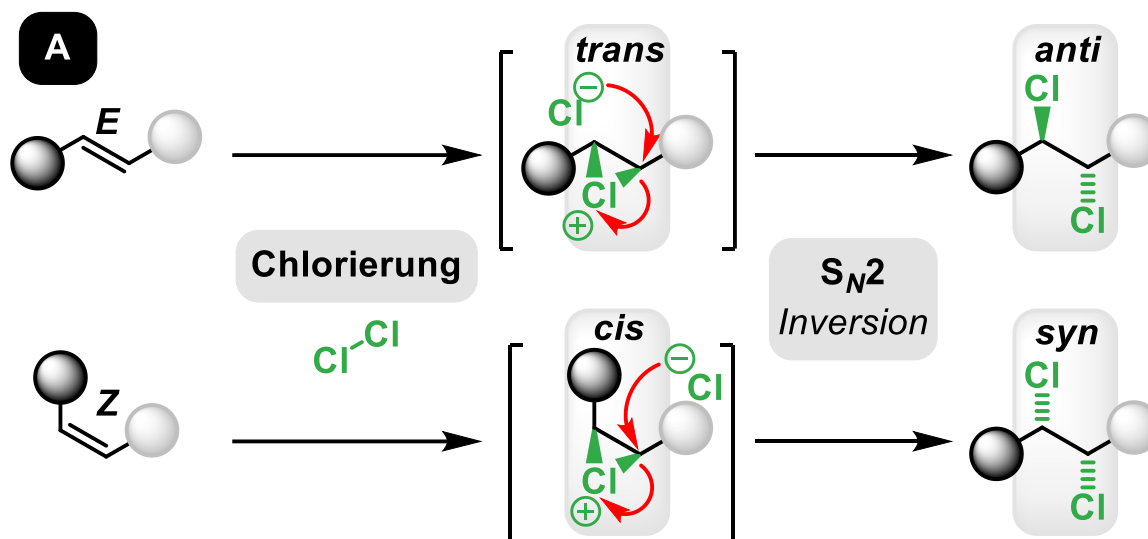
Mytilipin A **2001** (Y = Cl)

Deschloromytilipin A **2010** (Y = H)

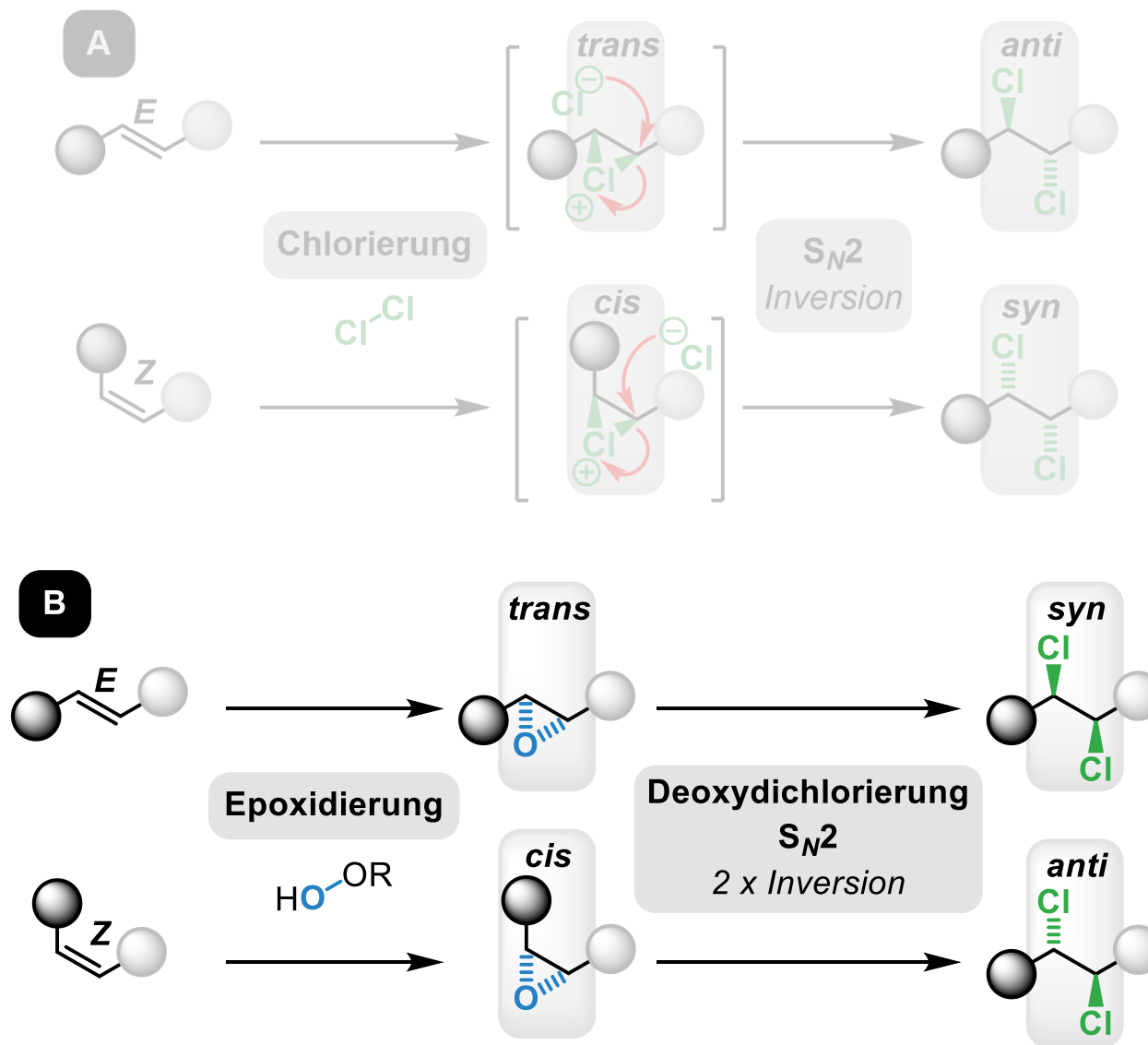


Mytilipin B **2002**
revidiert **2019**

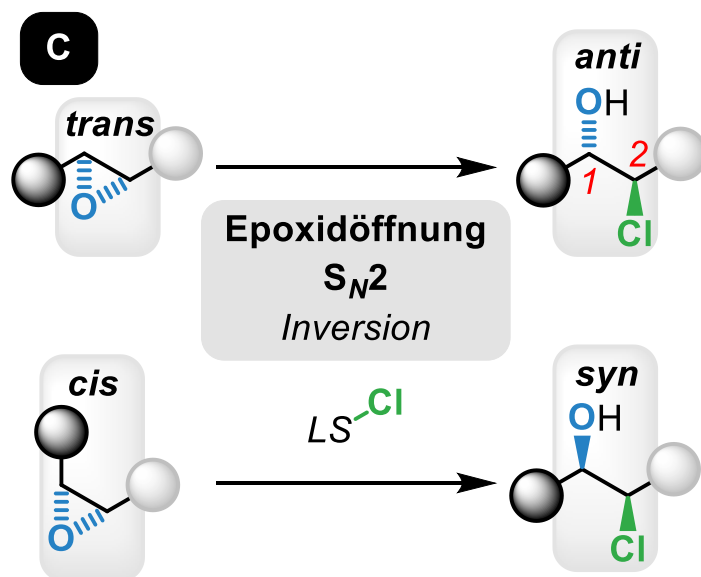
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Übersichtsartikel: (a) Vanderwal *et al.*, *Nat. Prod. Rep.* **2011**, 28, 15. (b) Carreira *et al.*, *Eur. J. Org. Chem.* **2012**, 1685. (c) Matsuda *et al.*, *Tetrahedron Lett.* **2014**, 55, 3003. (d) Vanderwal *et al.*, *Acc. Chem. Res.* **2014**, 47, 718. (e) P. H. Huy, P. Grewelinger, *Nachr. Chem.* **2019**, 67(9), 63.

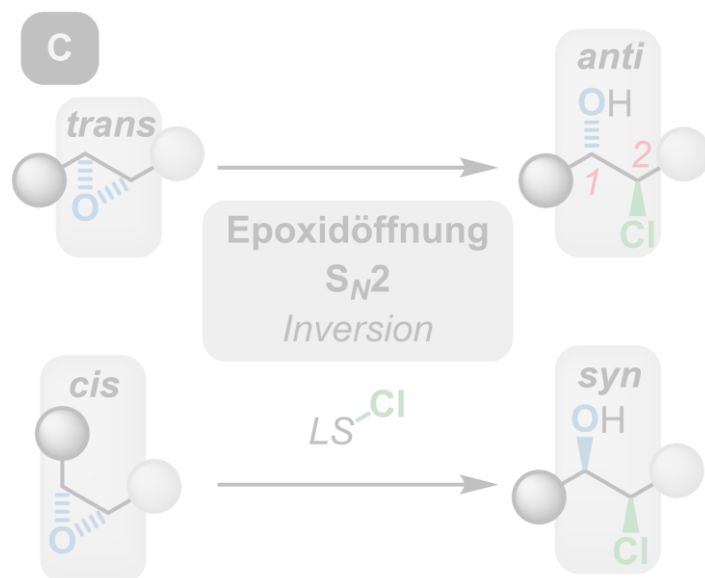


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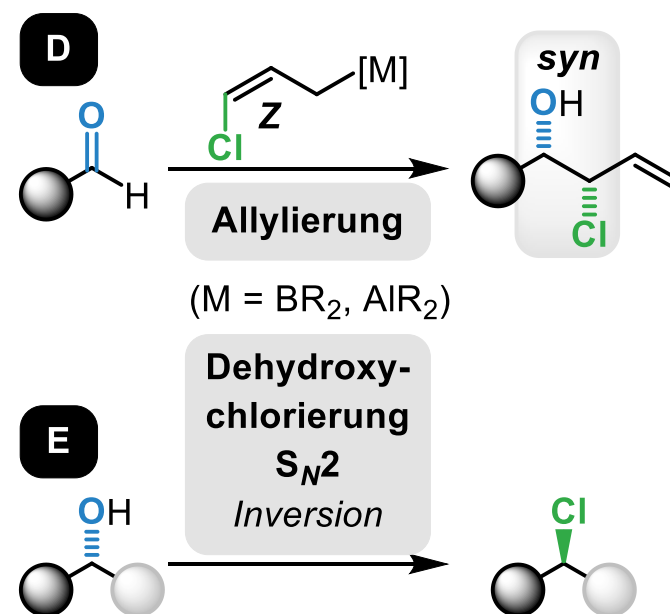
Lewis-Säuren (LS)

- Ti(+IV), Zr(+IV), B(+III), Si(+IV)

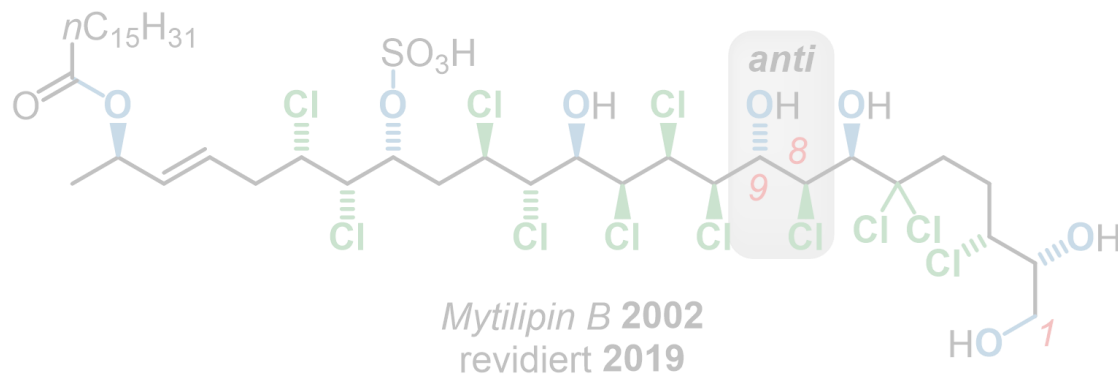
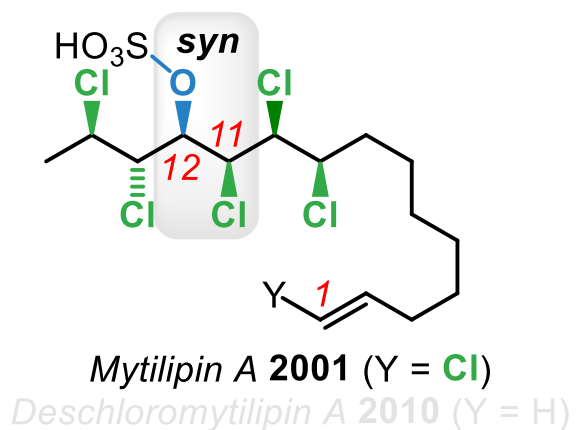
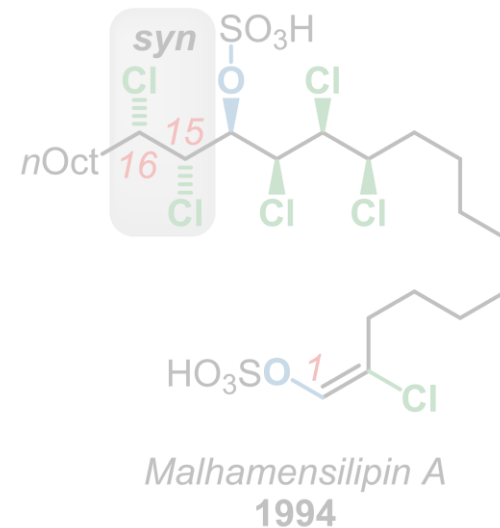
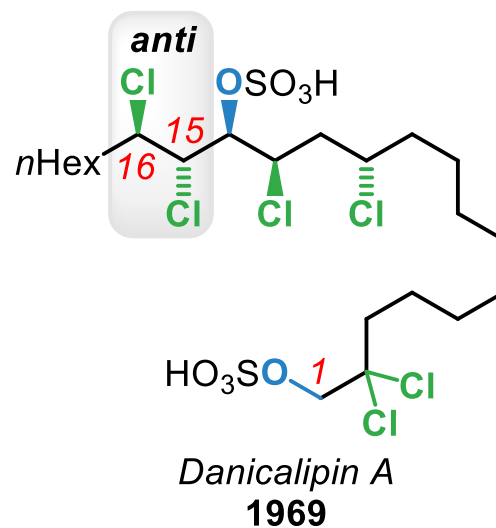


Lewis-Säuren (LS)

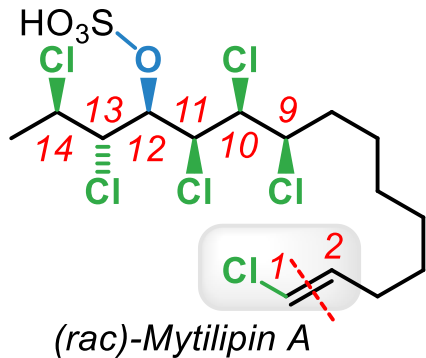
• Ti(+IV), Zr(+IV), B(+III), Si(+IV)



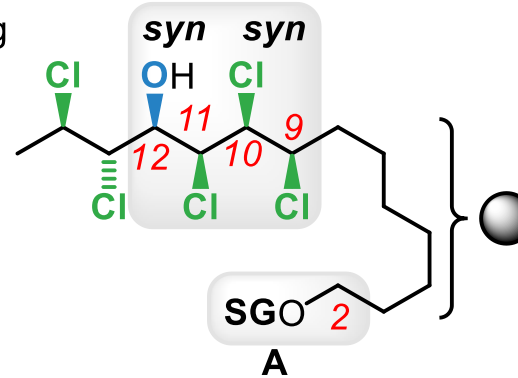
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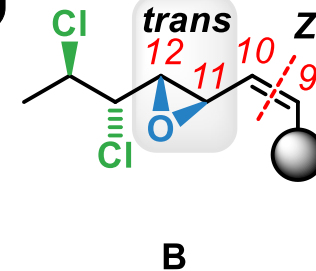
Carreira 2009



1. Sulfonierung
2. Takai-Alkenylierung
3. Oxidation
4. Entschützung

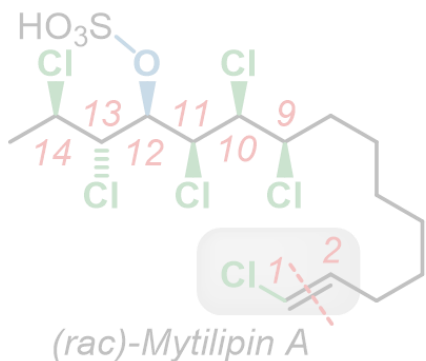


1. Chlorierung
2. Epoxidöffnung

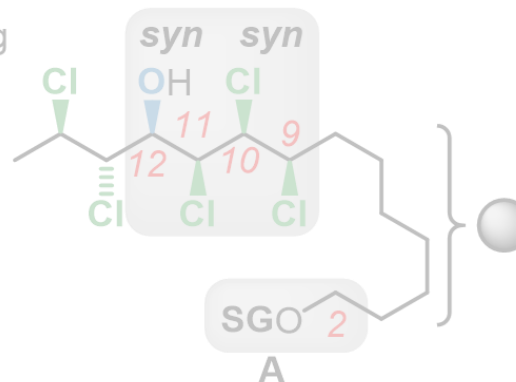


- Isoliert von Muschel *Mytilus galloprovincialis*
- 6 Stereozentren
- 11 lineare Stufen

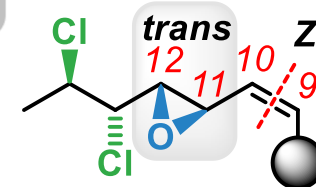
Carreira 2009



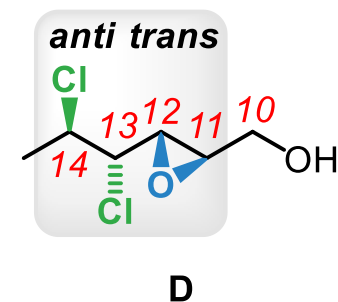
1. Sulfonierung
2. Takai-Alkenylierung
3. Oxidation
4. Entschützung



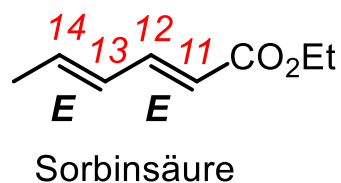
1. Chlorierung
2. Epoxidöffnung



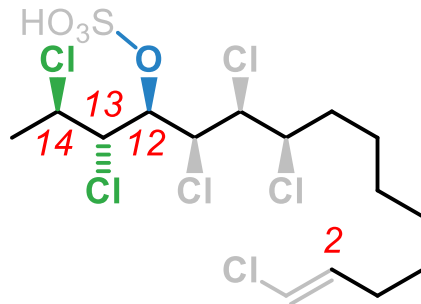
1. Wittig-Alkenylierung
2. Oxidation



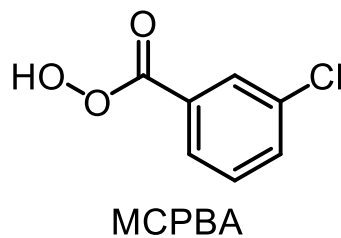
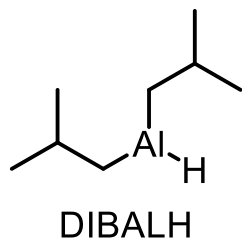
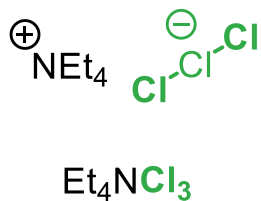
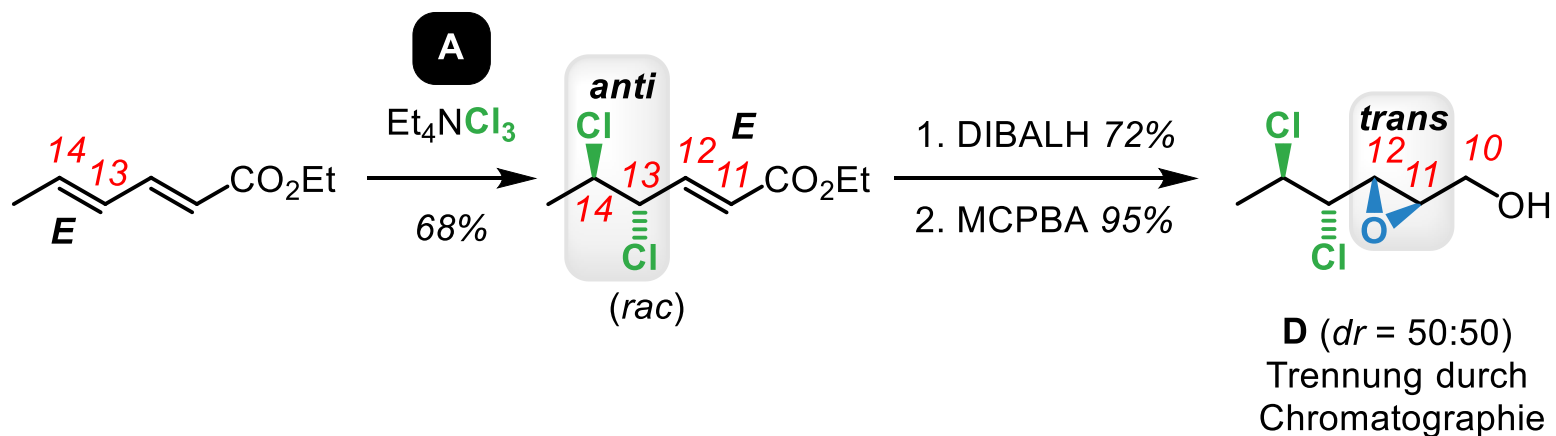
1. Epoxidierung
2. Reduktion
3. Chlorierung



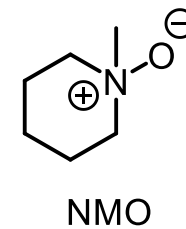
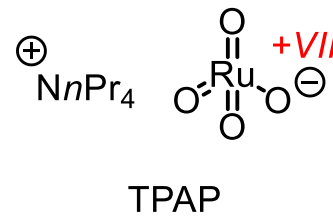
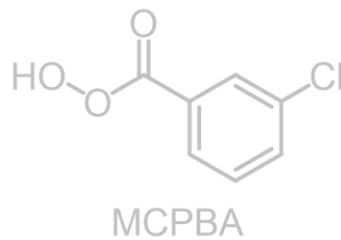
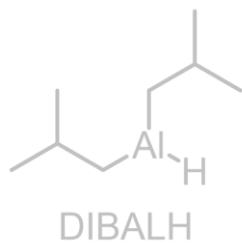
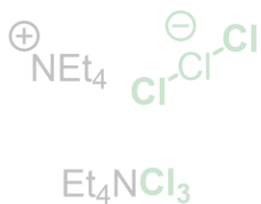
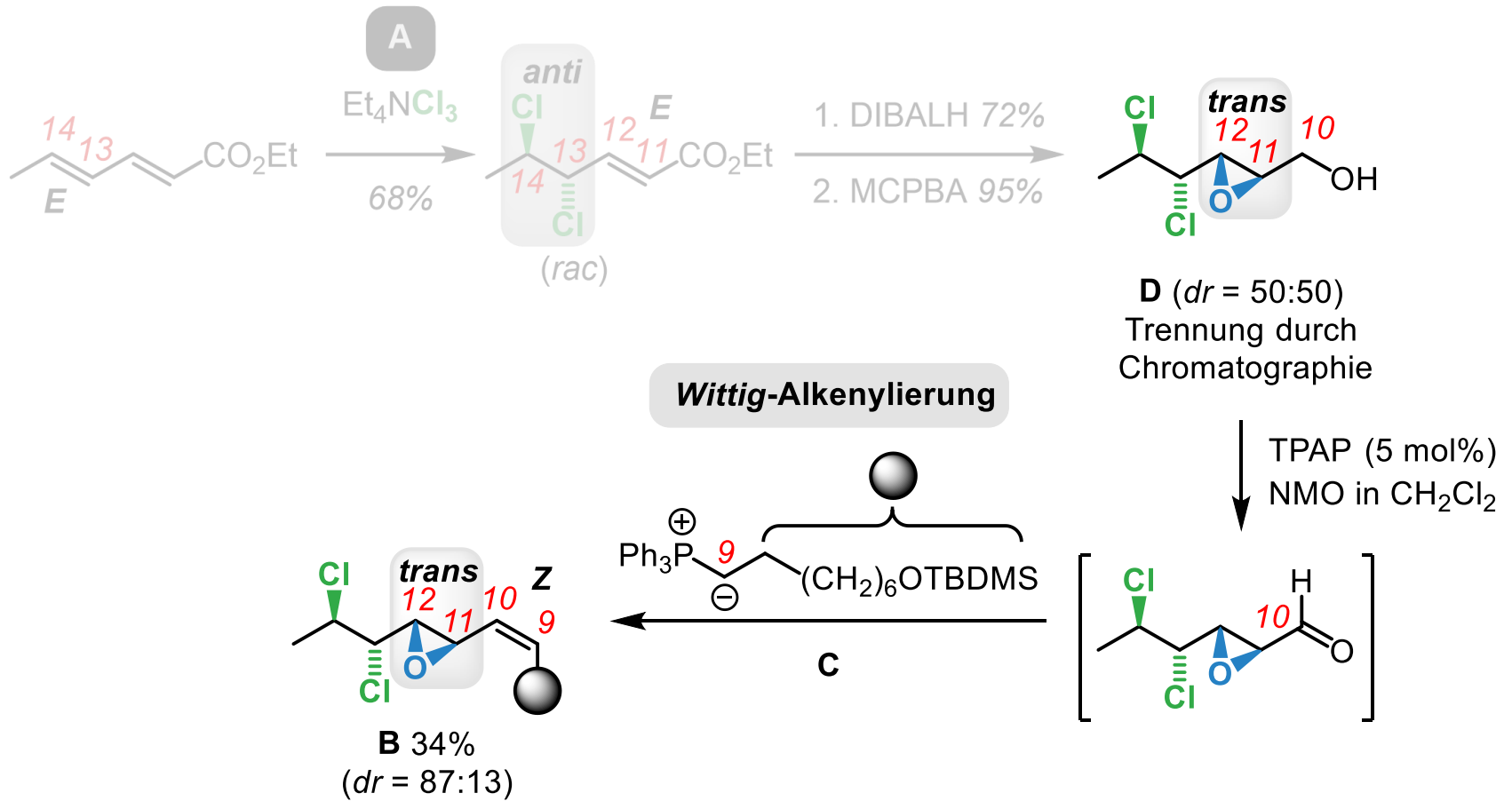
- Isoliert von Muschel *Mytilus galloprovincialis*
- 6 Stereozentren
- 11 lineare Stufen

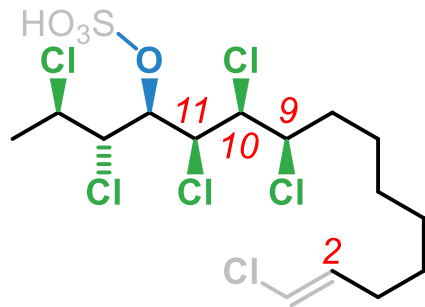


(rac)-Mytilipin A

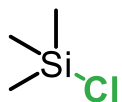
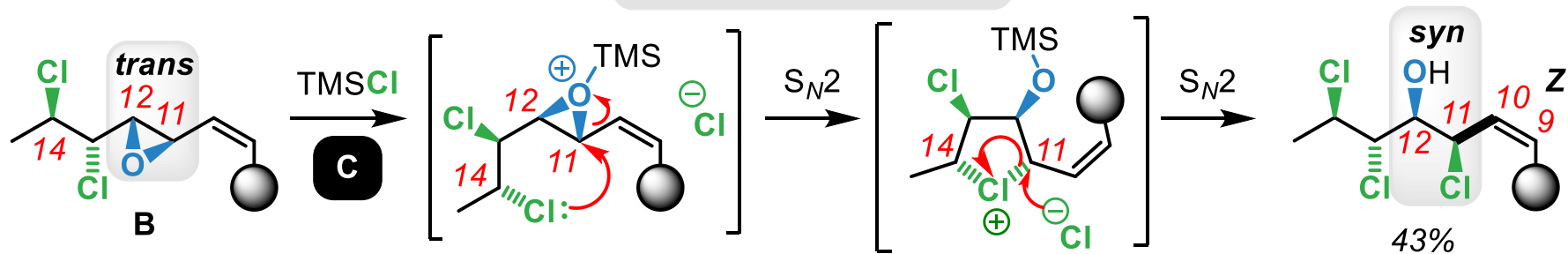


3. Synthese von Mytilipin A

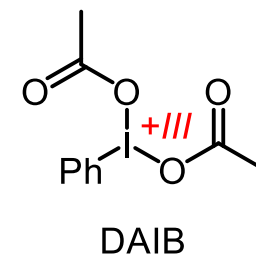
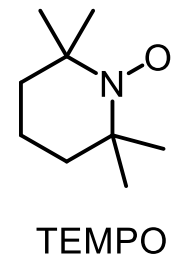
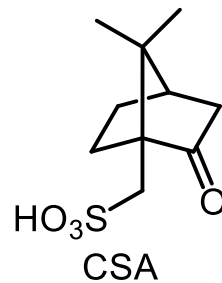
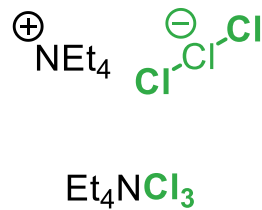
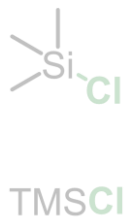
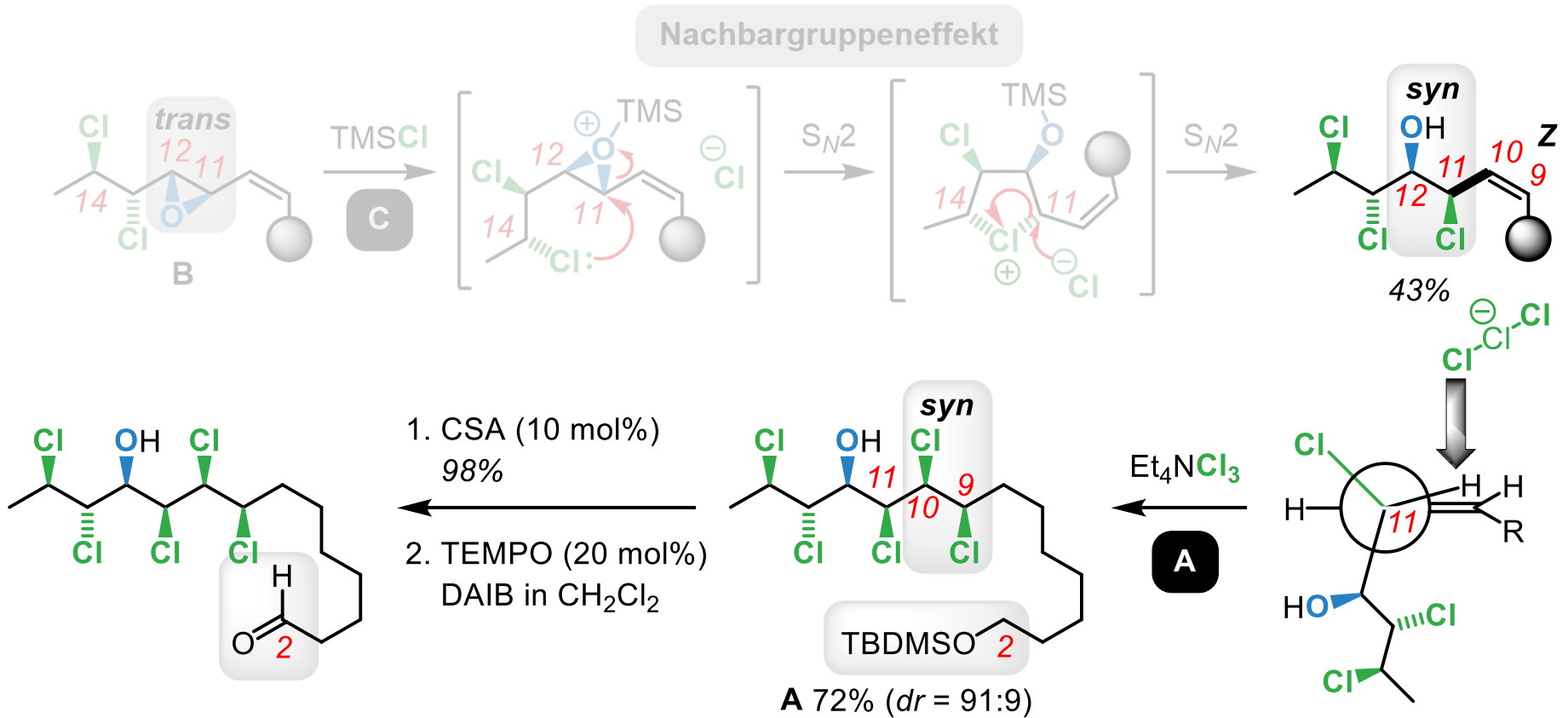


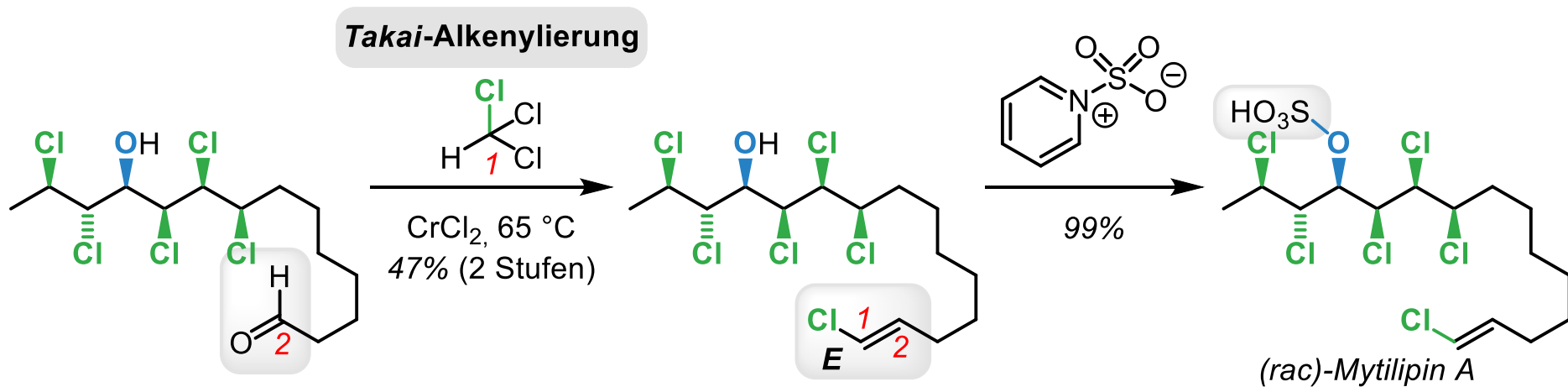


Nachbargruppeneffekt

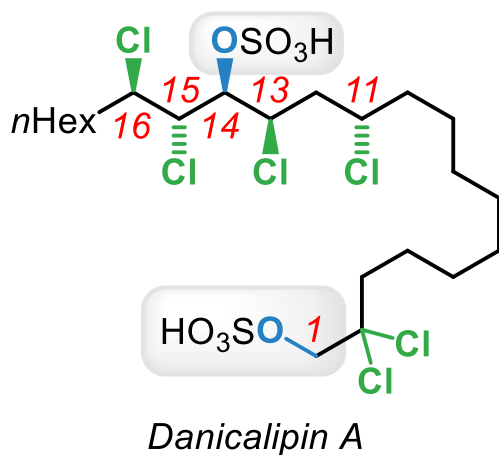


TMSCl



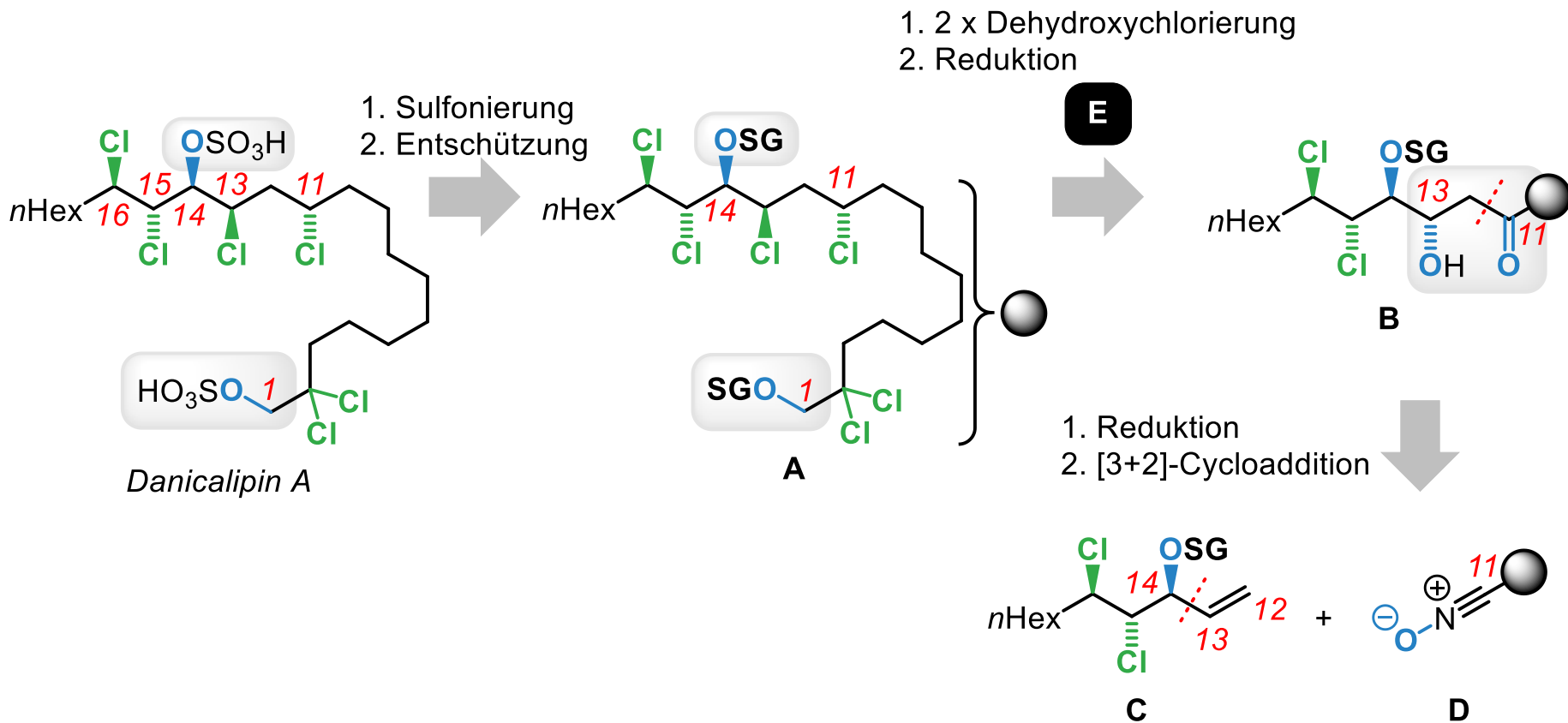


Yoshimitsu + Tanaka 2011

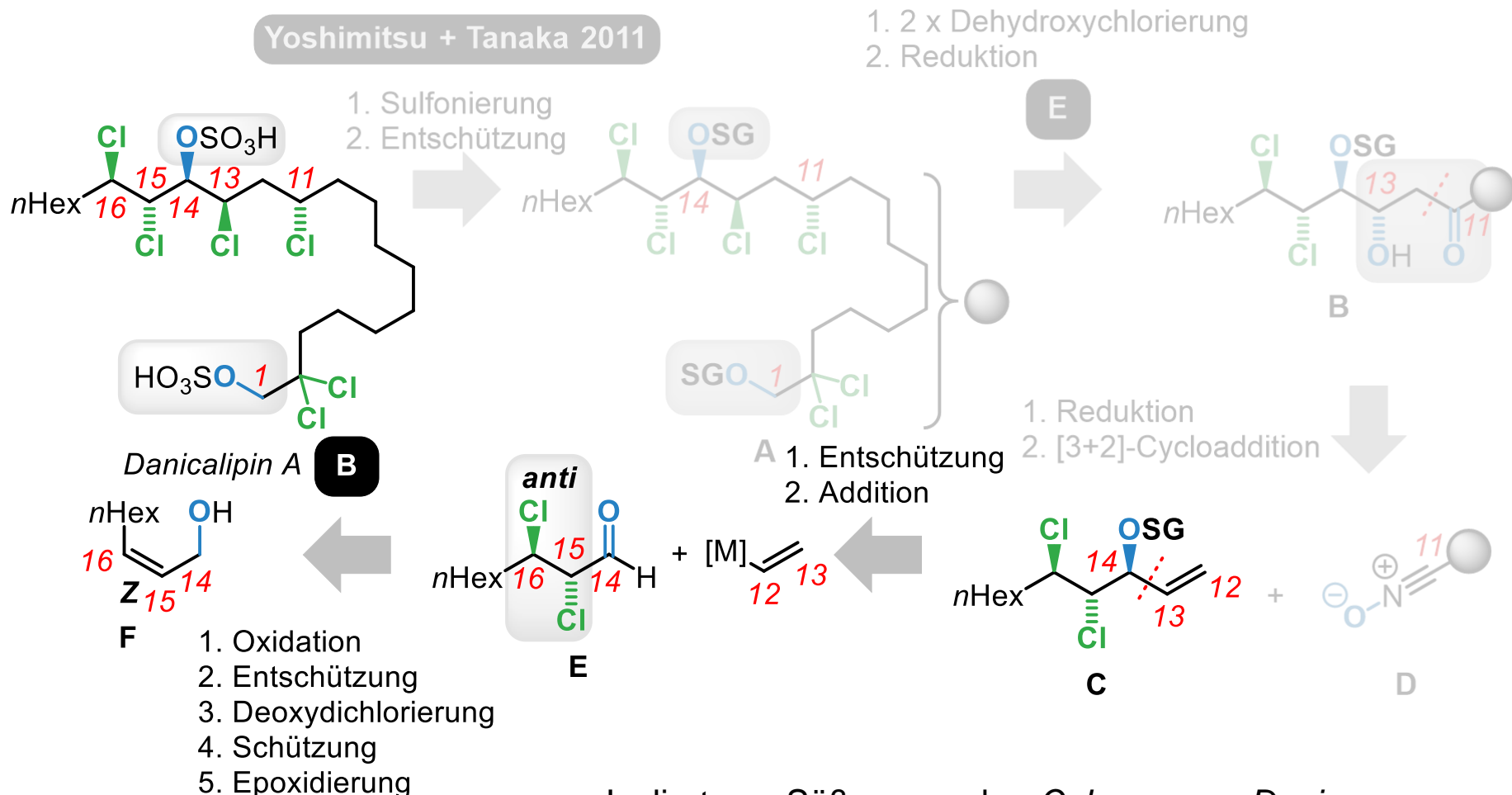


- Isoliert von Süßwasseralge *Ochromonas Danica*
- 5 Stereozentren
- 14 lineare Stufen

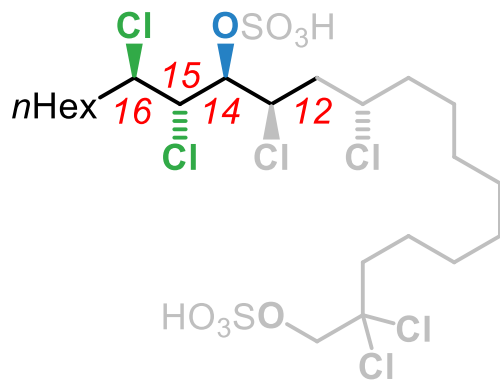
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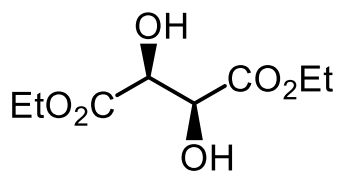
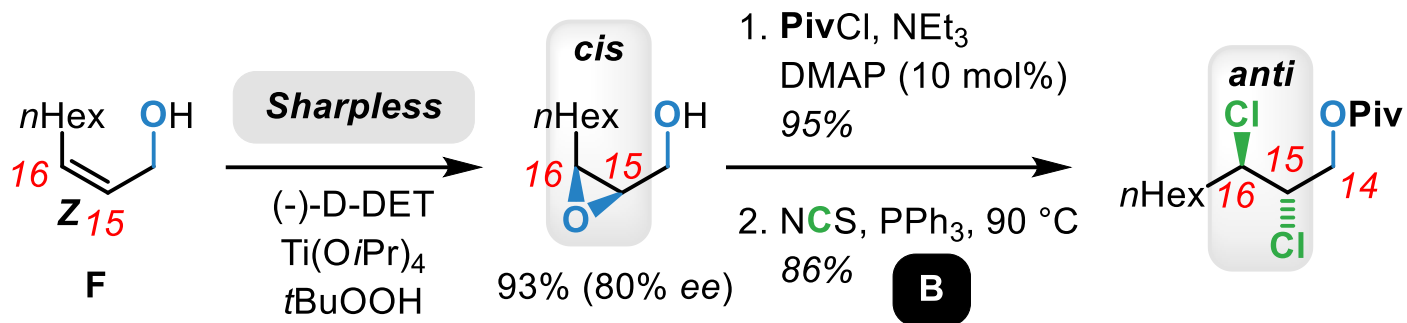


- Isoliert von Süßwasseralge *Ochromonas Danica*
- 5 Stereozentren
- 14 lineare Stufen

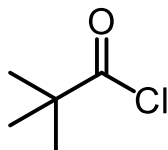


- Isoliert von Süßwasseralge *Ochromonas Danica*
- 5 Stereozentren
- 14 lineare Stufen

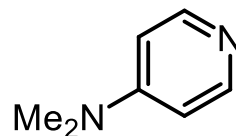




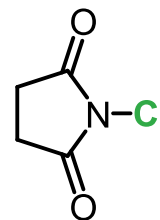
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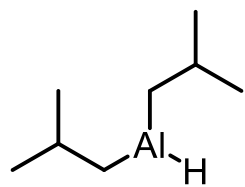
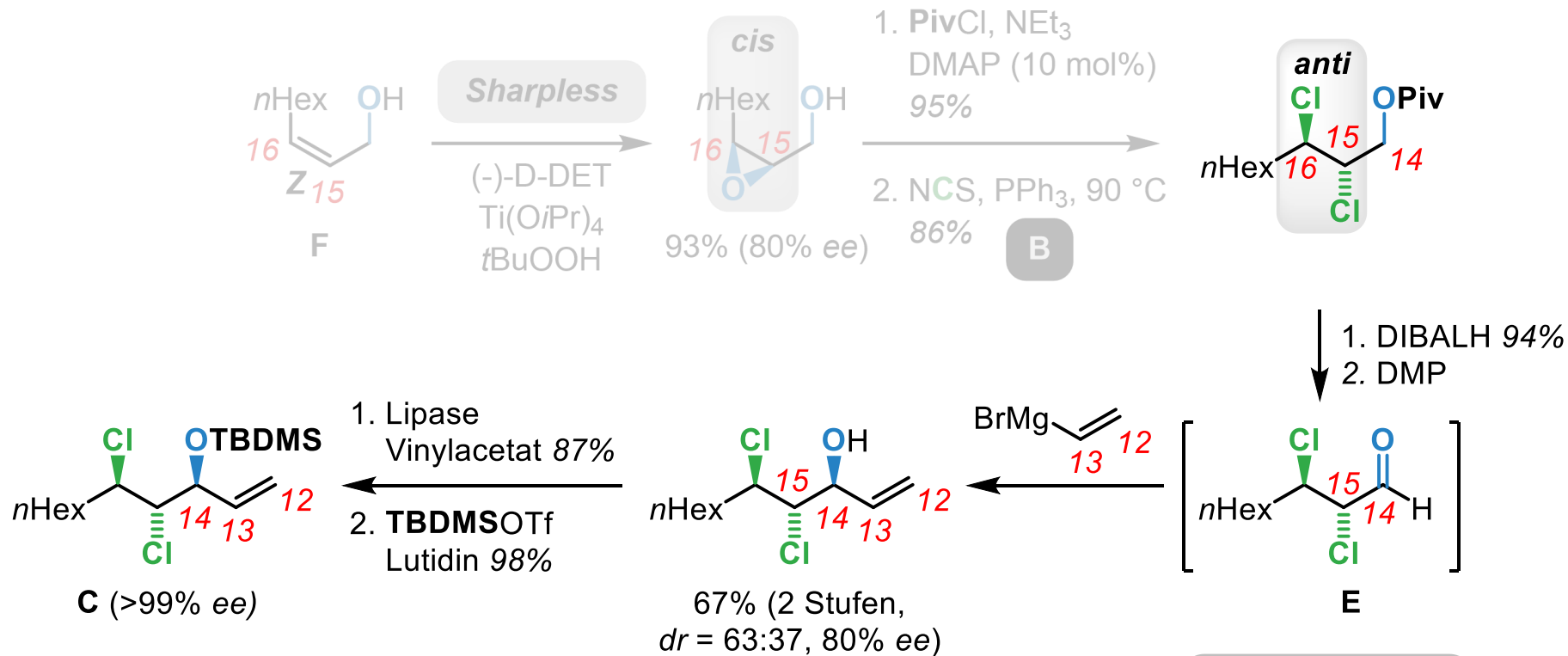
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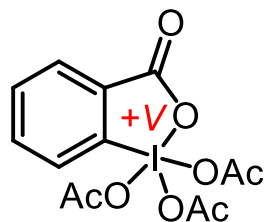
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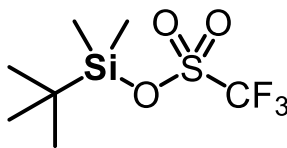
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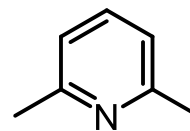
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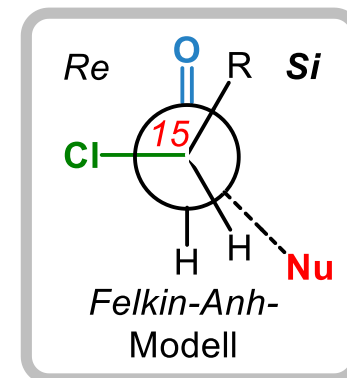
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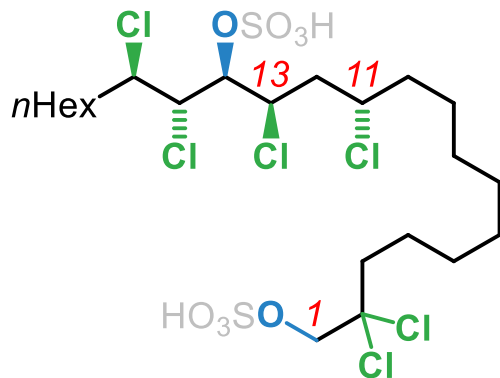


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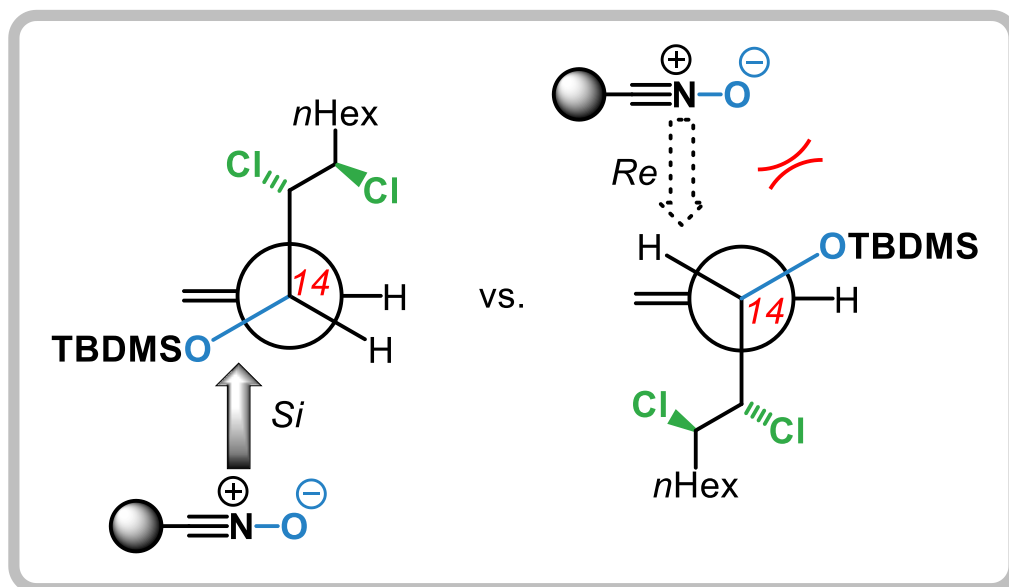
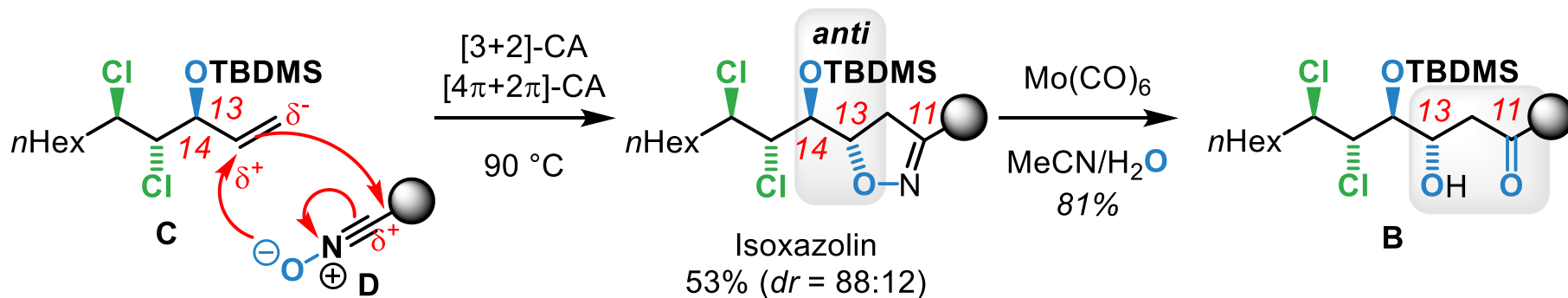


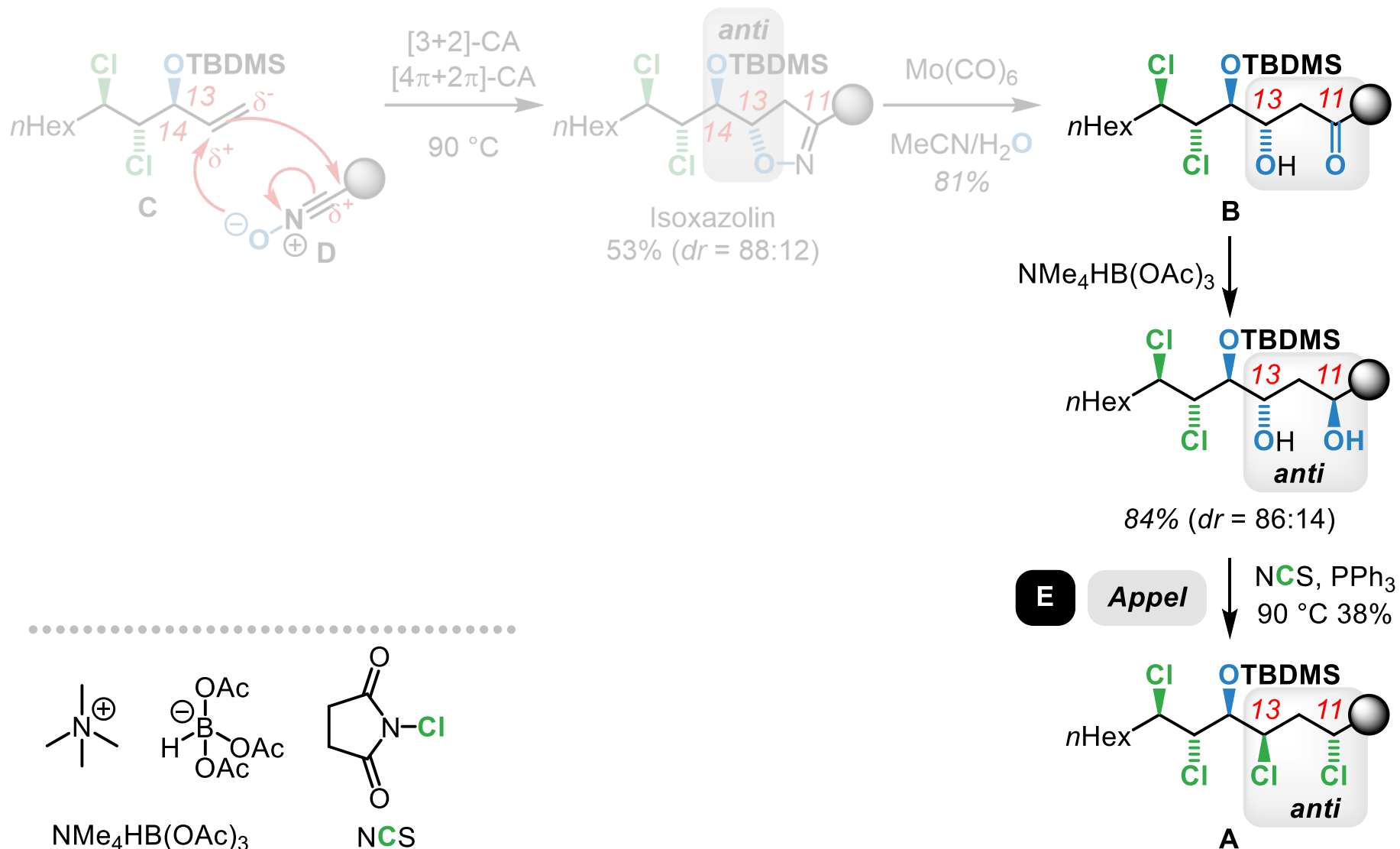
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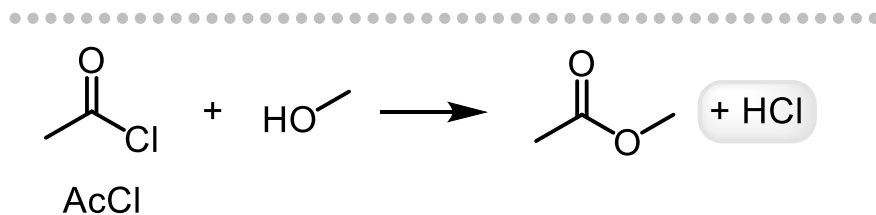
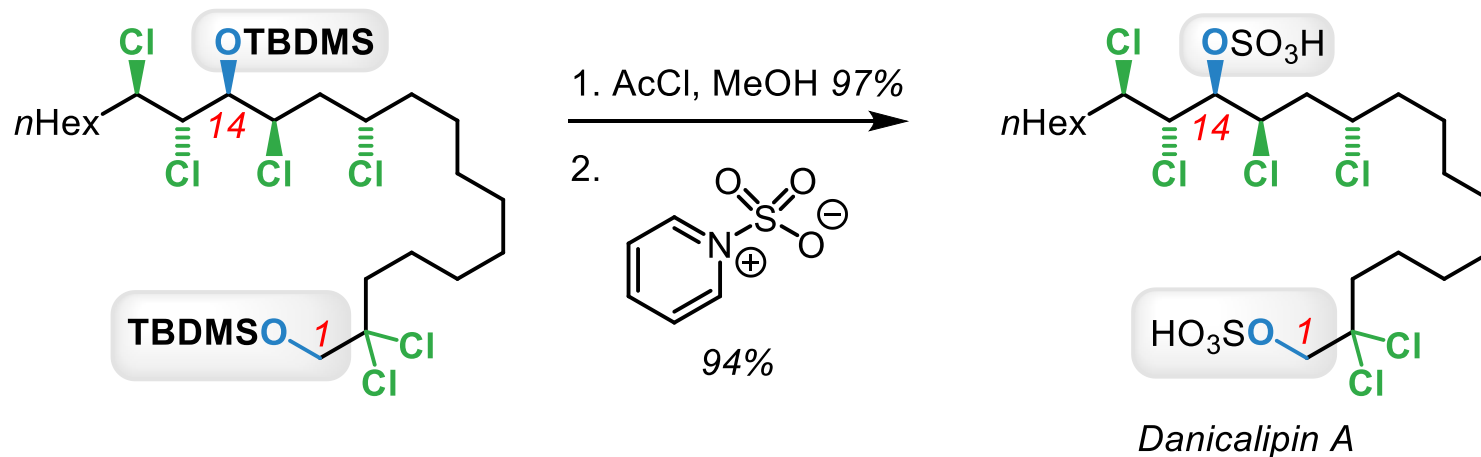


- (a) Houk *et al.*, *J. Am. Chem. Soc.* **1986**, 108, 2755. (b) Kobayashi *et al.*, *Chem. Soc. Perkin Trans. I* **1985**, 1401.
(c) Goti, Brandi *et al.*, *Tetrahedron Lett.* **1990**, 31, 3351. (d) Evans *et al.*, *J. Am. Chem. Soc.* **1988**, 110, 3560.

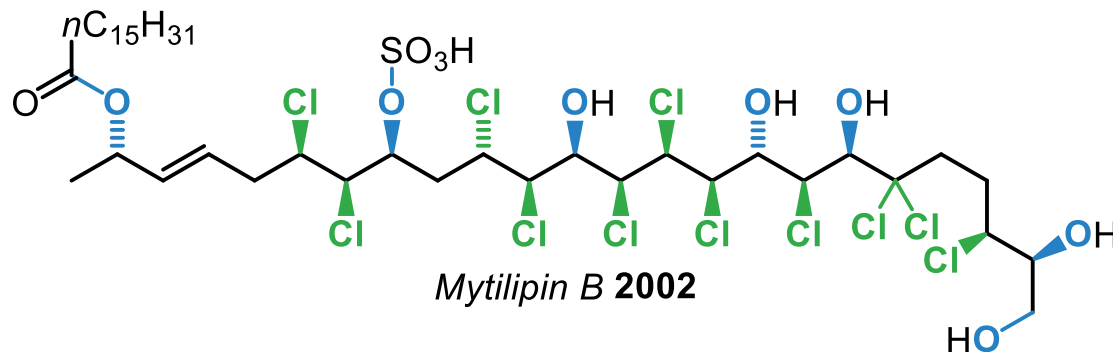




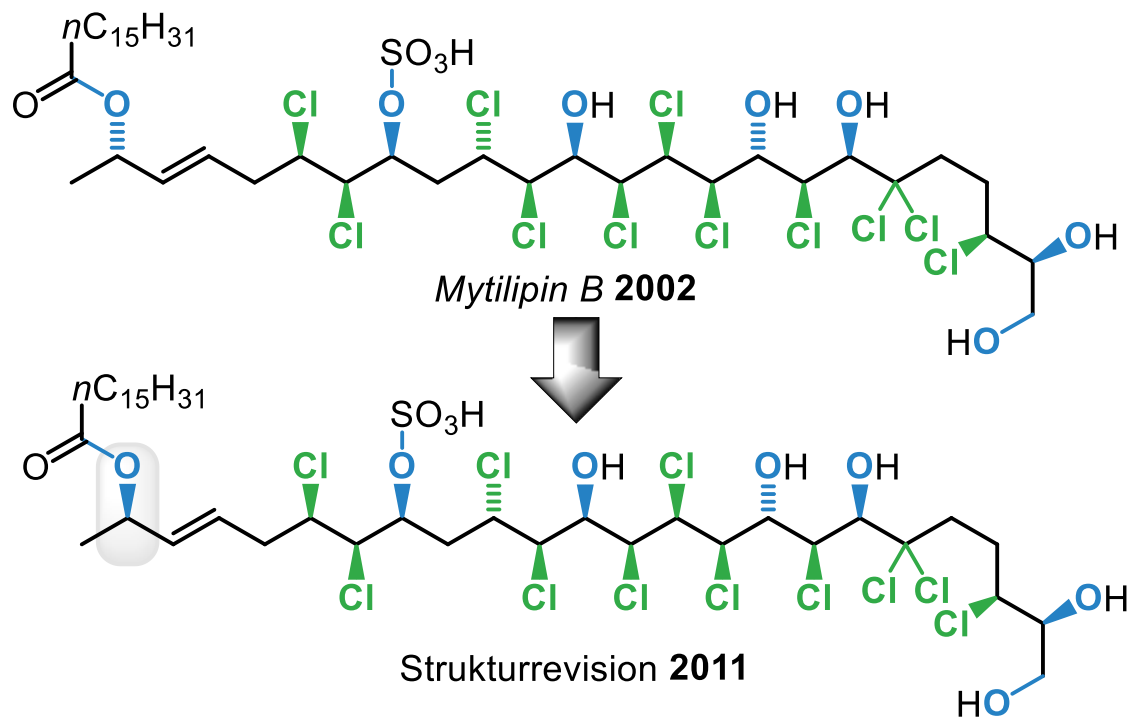
(a) Houk et al., *J. Am. Chem. Soc.* **1986**, 108, 2755. (b) Kobayashi et al., *Chem. Soc. Perkin Trans. I* **1985**, 1401.
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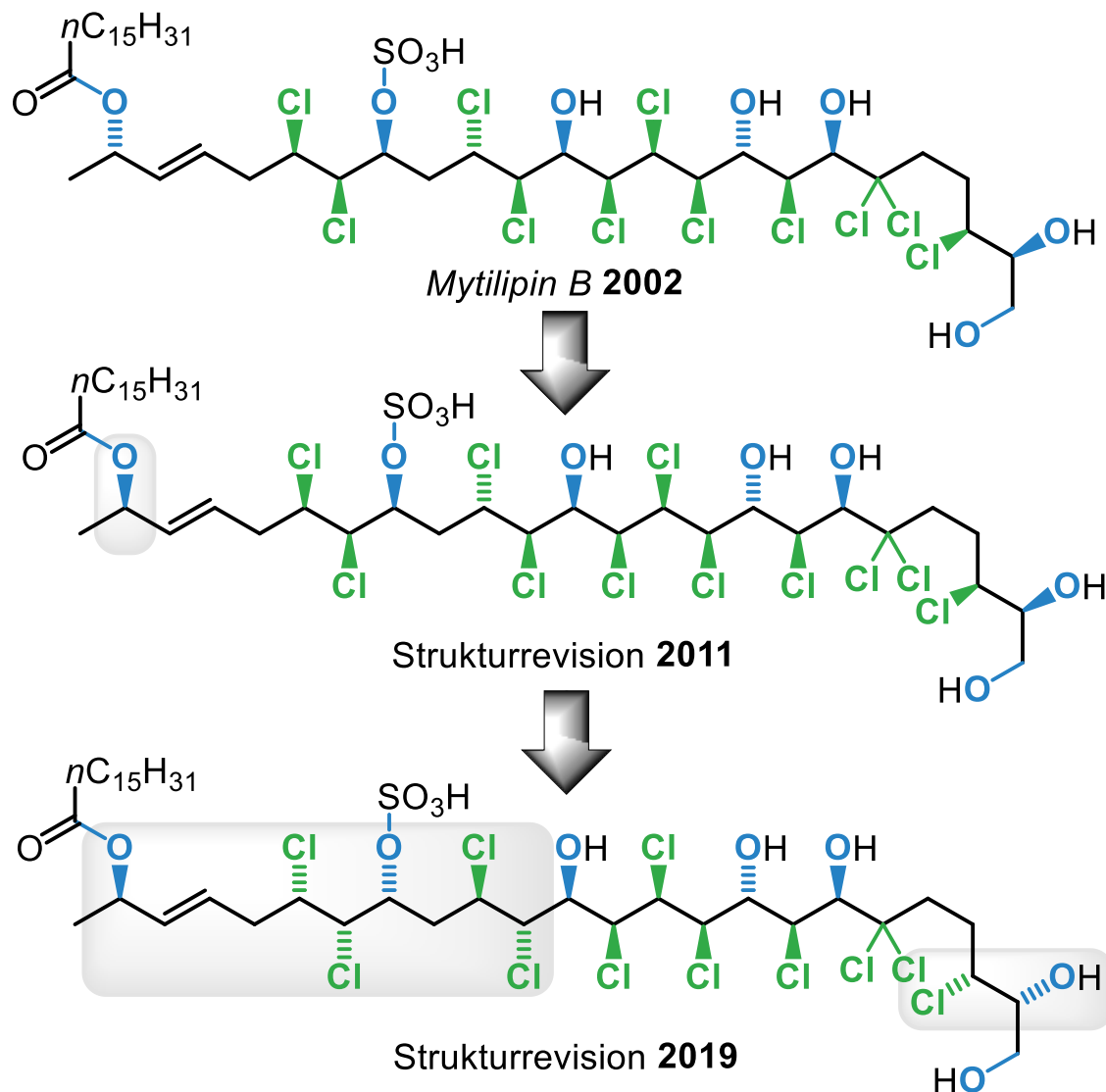
- Isoliert von Muschel
Mytilus galloprovincialis
- 15 Stereozentren



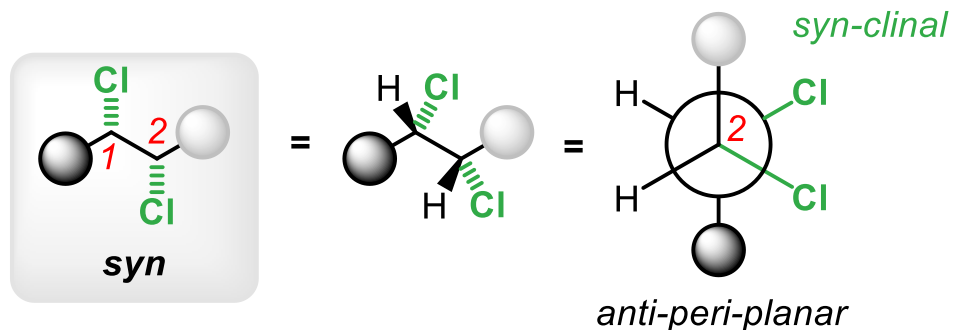
- Isoliert von Muschel
Mytilus galloprovincialis
- 15 Stereozentren



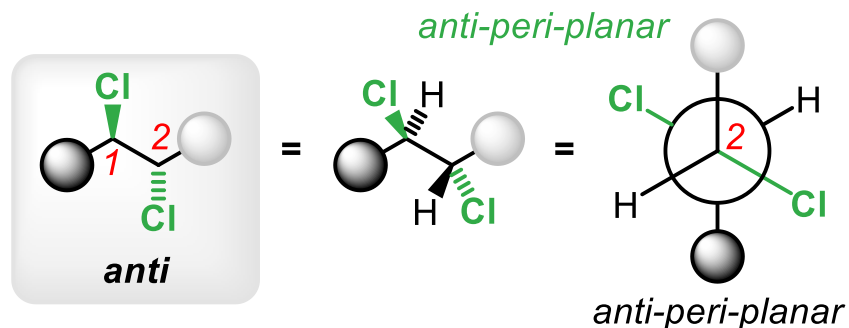
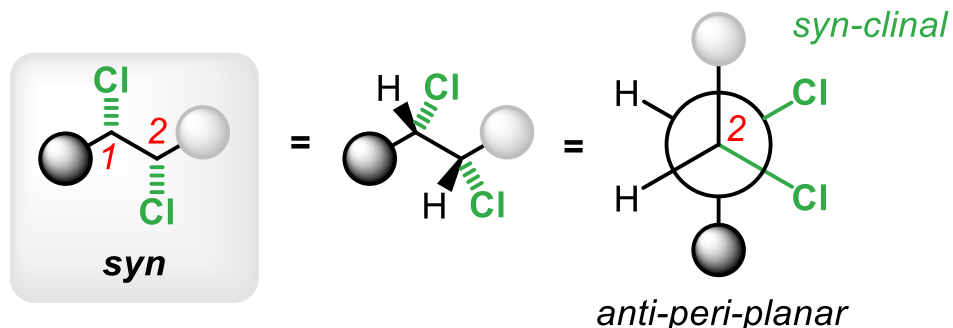
- Isoliert von Muschel
Mytilus galloprovincialis
- 15 Stereozentren
- 31 lineare Stufen



- Rigide Konformation
 - *Carreiras* Spektroskopische Datenbank
- Bestimmung von Konfiguration durch NMR-Kopplungskonstanten

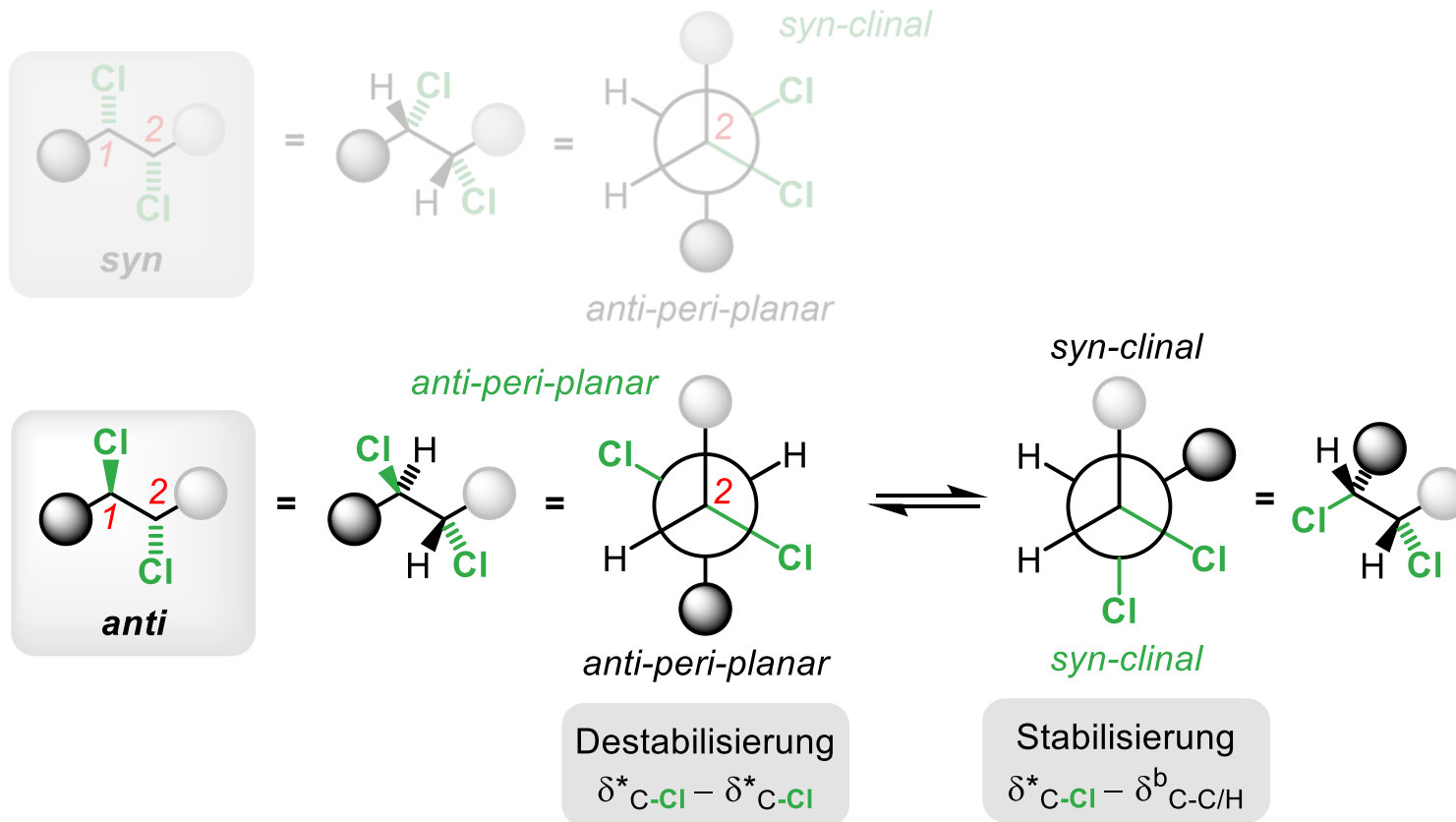


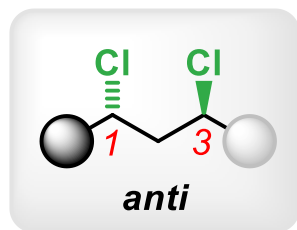
- Rigide Konformation
- Carreiras Spektroskopische Datenbank
- Bestimmung von Konfiguration durch NMR-Kopplungskonstanten



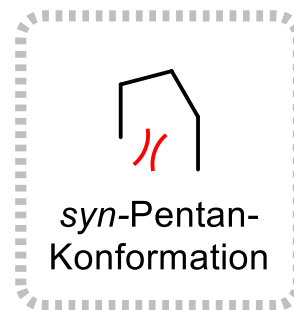
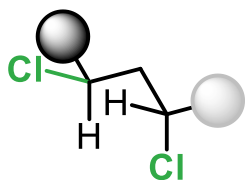
Destabilisierung
 $\delta^*_{\text{C-Cl}} - \delta^*_{\text{C-Cl}}$

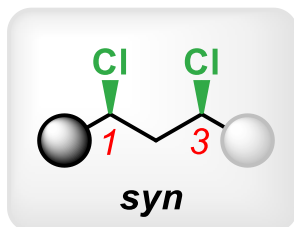
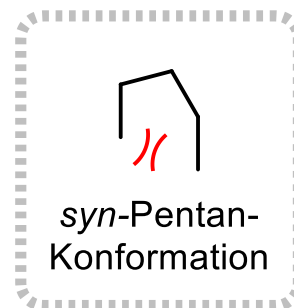
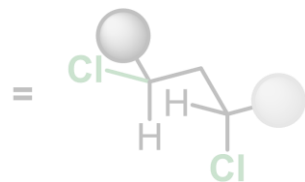
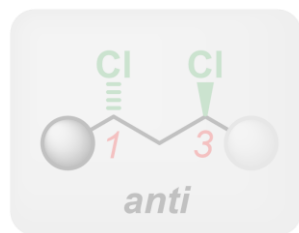
- Rigide Konformation
- Carreiras Spektroskopische Datenbank
- Bestimmung von Konfiguration durch NMR-Kopplungskonstanten



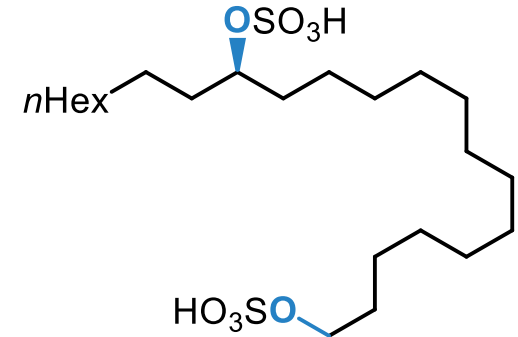
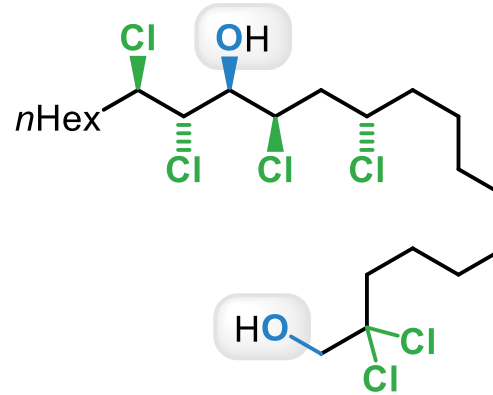
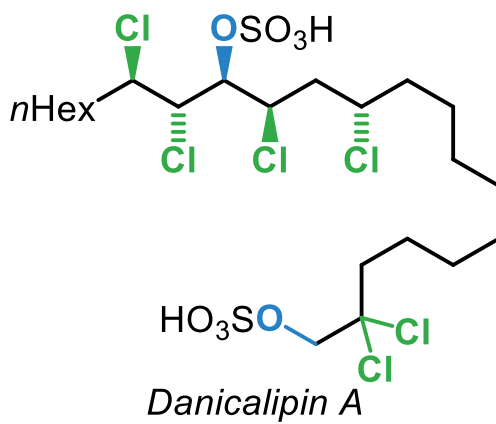


=





Toxizität
 <30 μM
 30-100 μM
 >100 μM

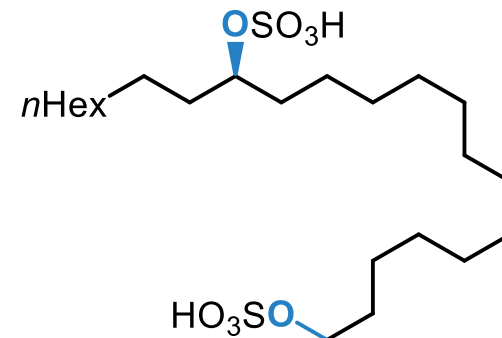
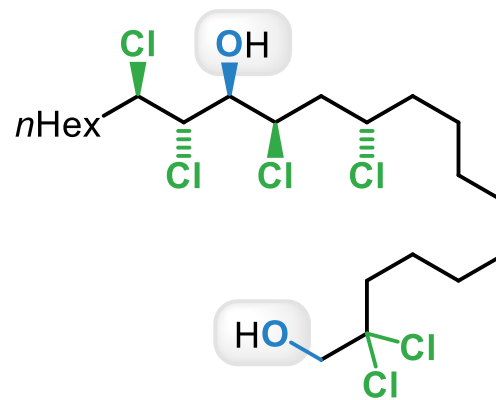
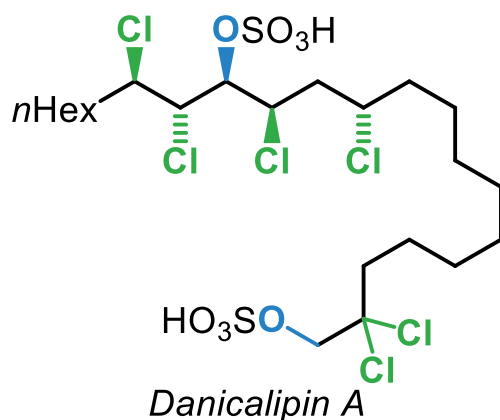


Salzwassergarnele **LC**₅₀ [μM] **5.3**
 A549 Zellen **EC**₅₀ [μM] **26.5**
 HT-29 Zellen **EC**₅₀ [μM] **15.5**
 Hepa 1-6 Zellen **EC**₅₀ [μM] **14.3**

>141
41.4
>166
17.3

63.8
69.3
84.4
39.1

Toxizität
 <30 μM
 30-100 μM
 >100 μM

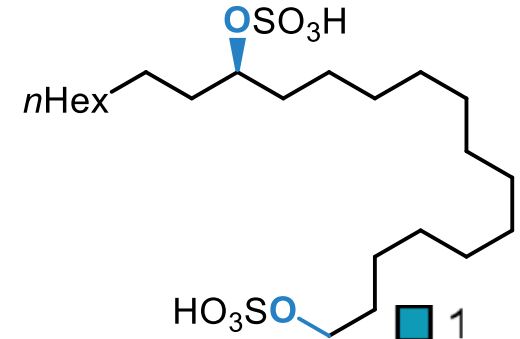
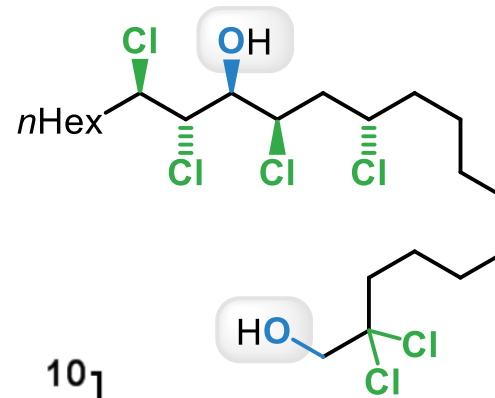
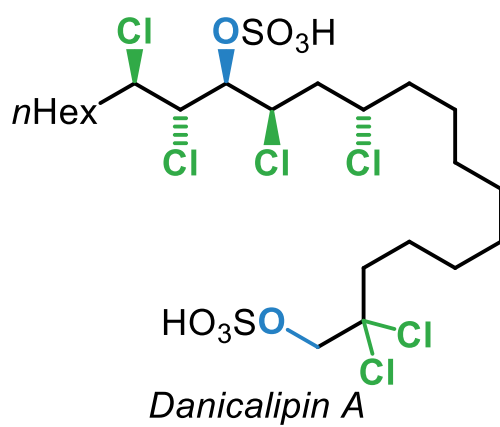


Salzwassergarnele **LC₅₀** [μM] **5.3**
 A549 Zellen **EC₅₀** [μM] **26.5**
 HT-29 Zellen **EC₅₀** [μM] **15.5**
 Hepa 1-6 Zellen **EC₅₀** [μM] **14.3**

>141
41.4
>166
17.3

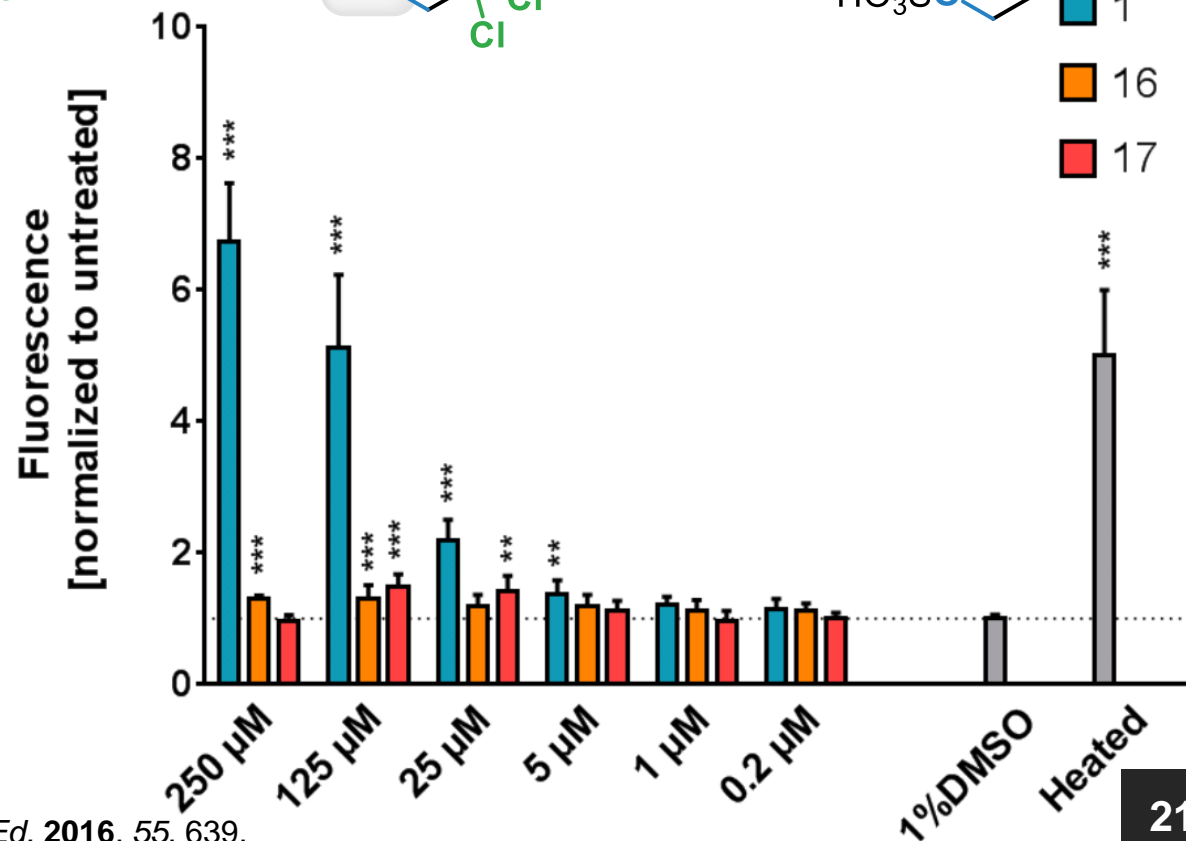
63.8
69.3
84.4
39.1

→ Toxizität : **Cl-Atom** und **Sulfonatgruppen** essentiell

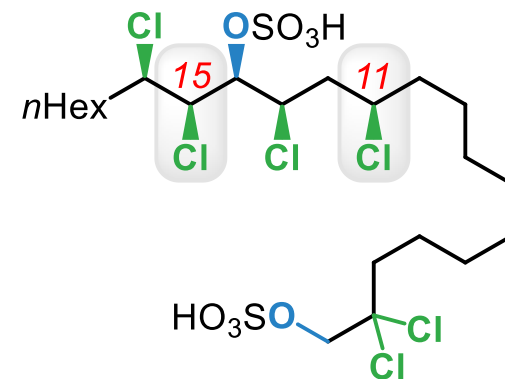
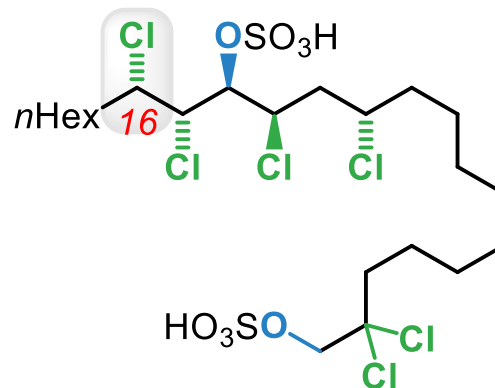
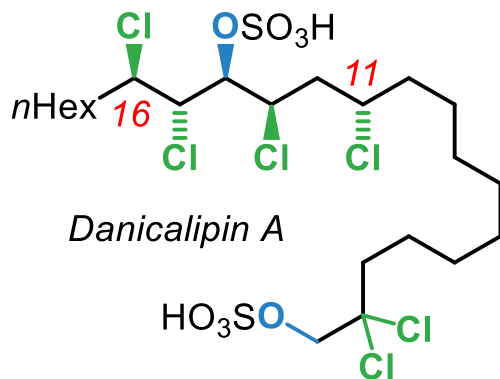


Permeabilität Zellmembran

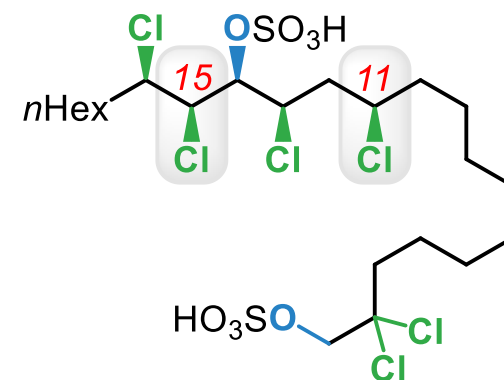
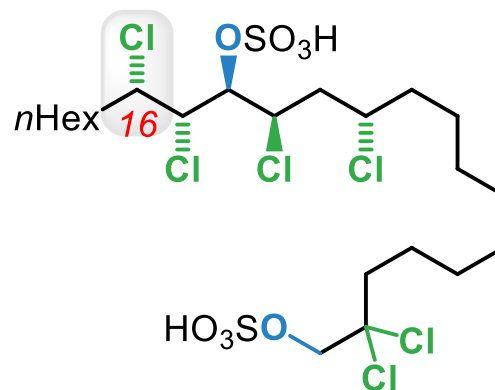
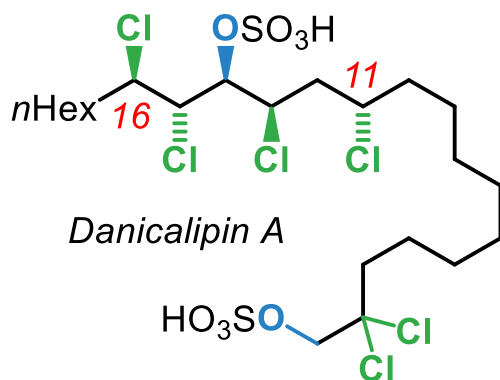
Fluoreszenz von Gram-negativen *E. coli* DH5 α Bakterien mittels Hoechst 33342



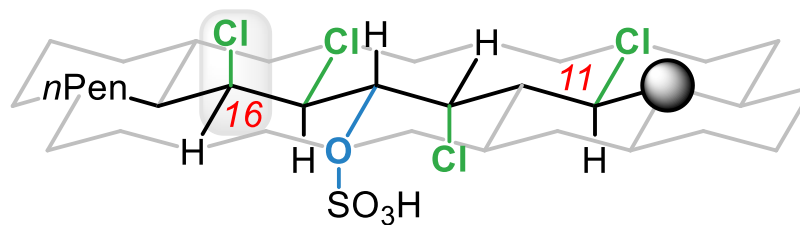
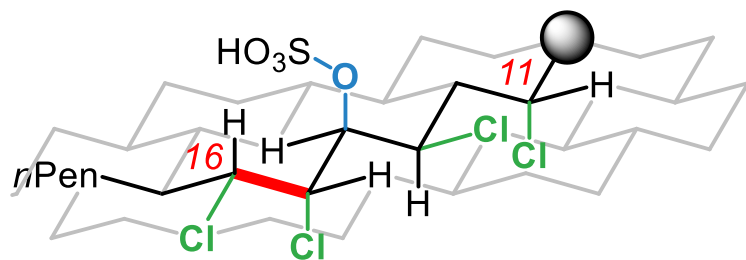
Toxizität

<5 μM 5-20 μM >20 μM 

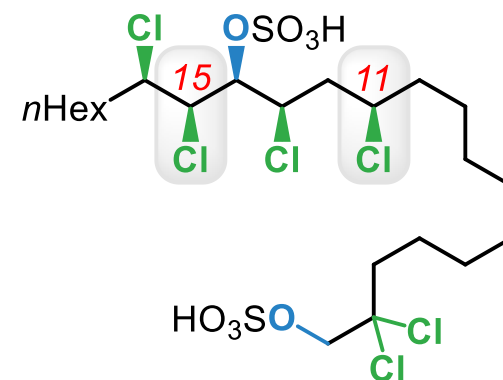
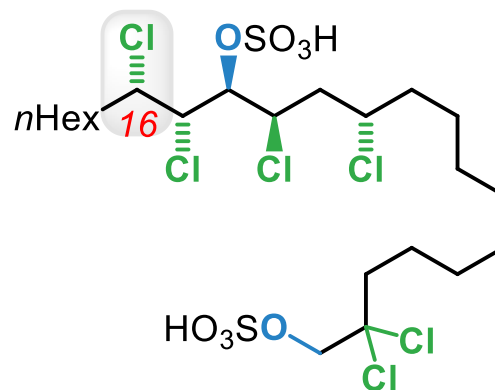
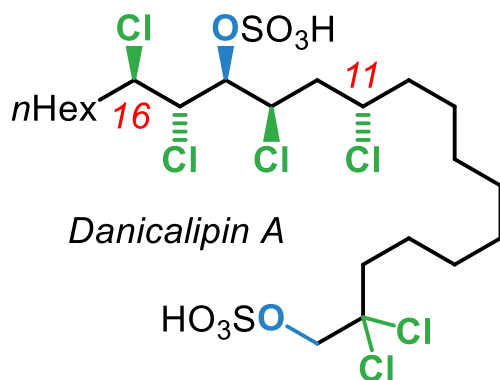
Toxizität

<5 μM 5-20 μM >20 μM 

Änderung Vorzugskonformation



Toxizität
 <5 μM
 5-20 μM
 >20 μM

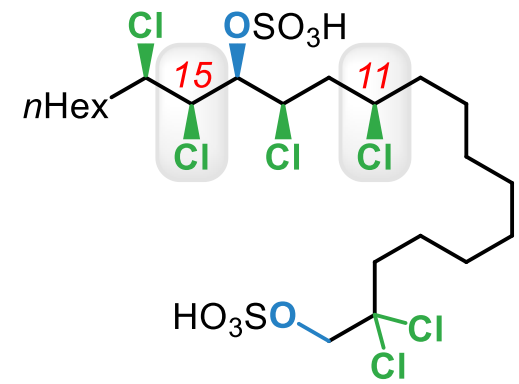
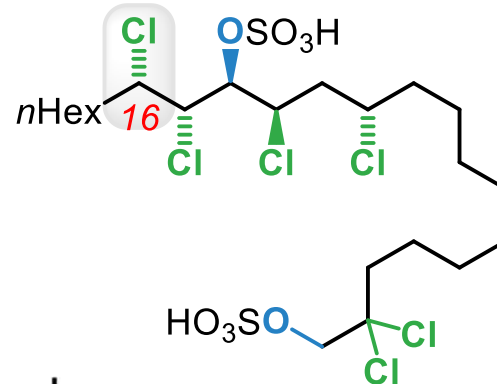
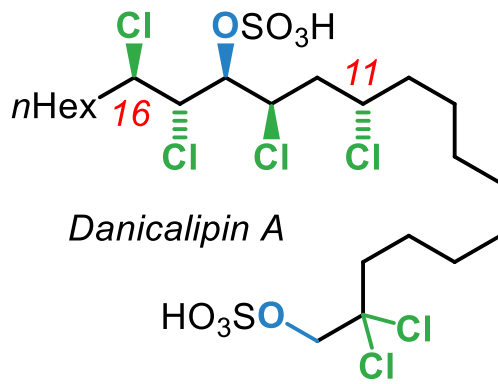


Salzwassergarnele **LC**₅₀ [μM] **2.5**
 A549 Zellen **EC**₅₀ [μM] **32.2**
 HT-29 Zellen **EC**₅₀ [μM] **14.7**
 Hepa 1-6 Zellen **EC**₅₀ [μM] **14.7**

5.7
42.6
10.9
13.3

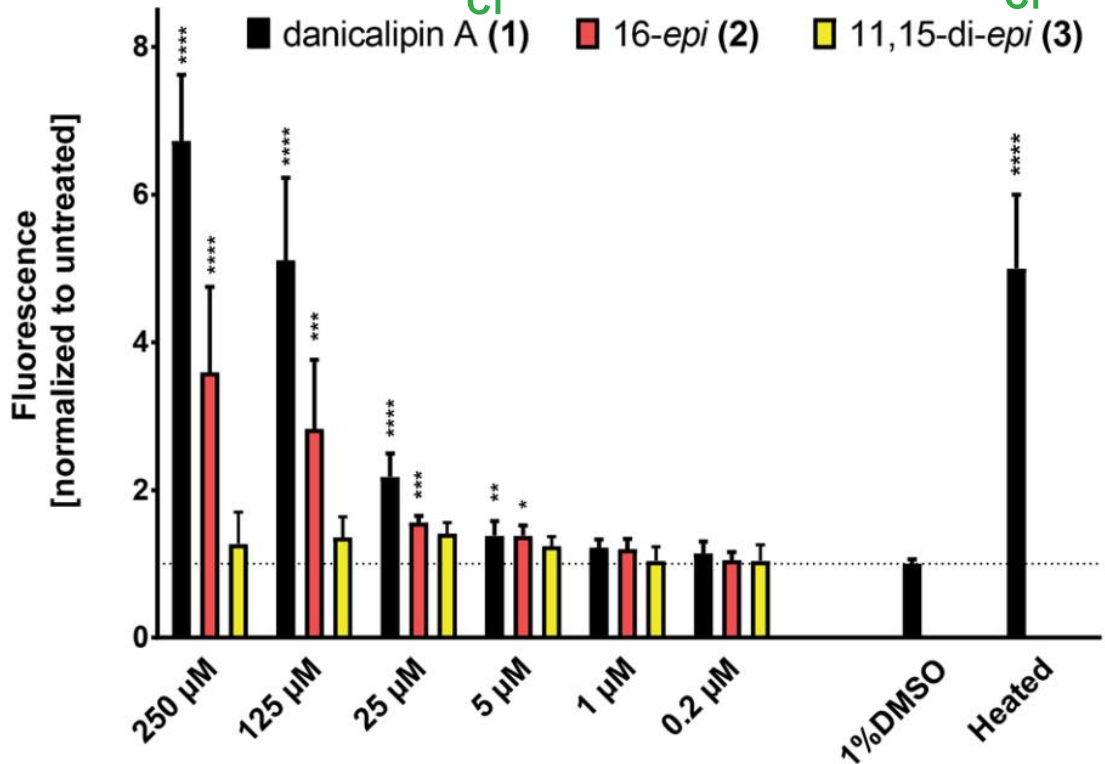
4.5
36.0
3.7
10.2

→ Toxizität: *Einfluss relative Konfiguration gering*



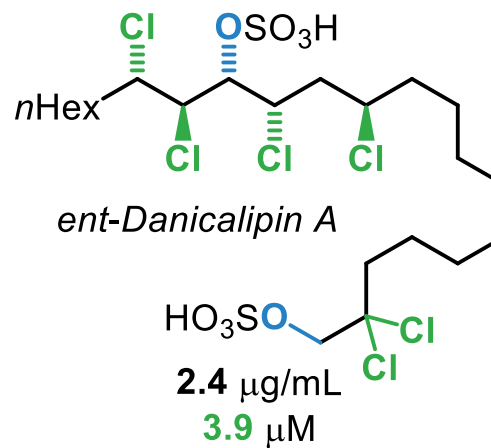
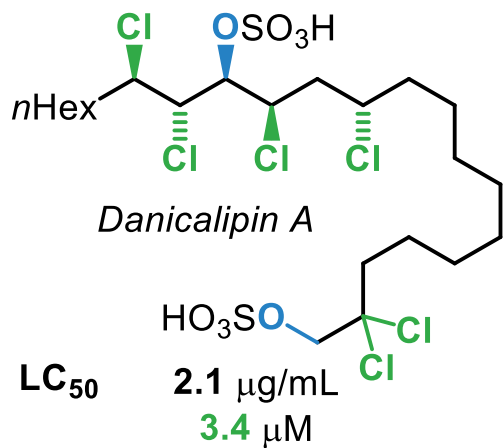
Permeabilität Zellmembran

Fluoreszenz von Gram-negativen *E. coli* DH5 α Bakterien mittels Hoechst 33342



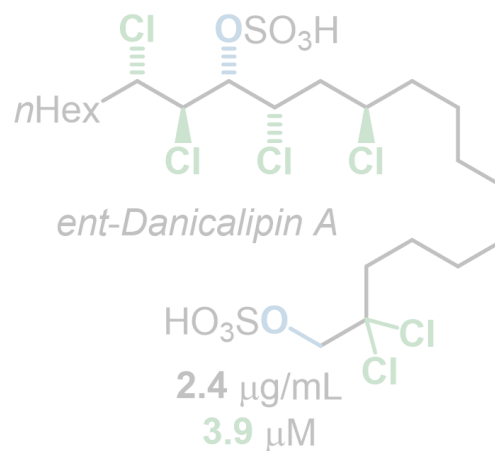
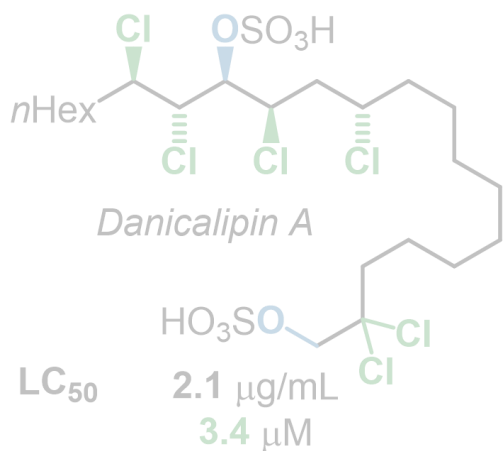
→ Zellpermeabilität: **Relative Konfiguration** essentiell

Toxizität
<30 μM
30-100 μM
>100 μM

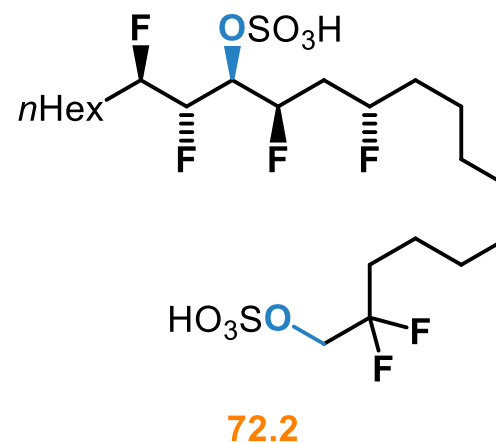
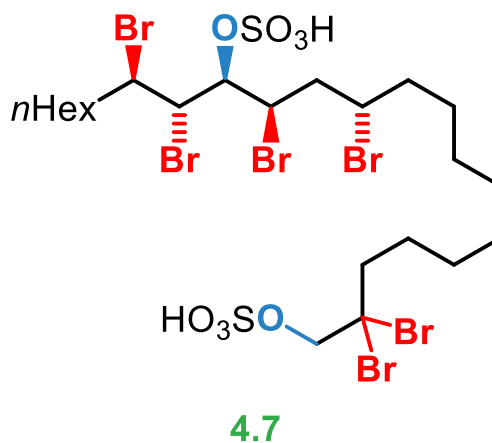
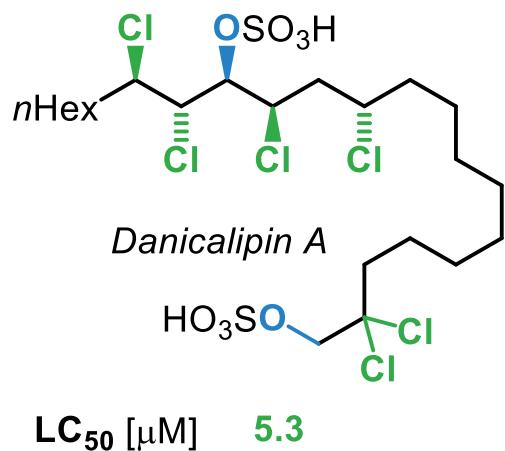


→ Toxizität Salzwassergarnelen: Absolute Konfiguration keinen Einfluss

Toxizität
 <30 μM
 30-100 μM
 >100 μM

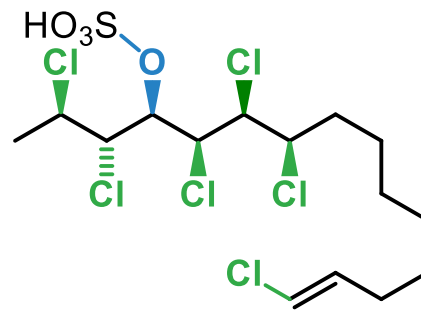


→ Toxizität Salzwassergarnelen: Einfluss Absolute Konfiguration keinen

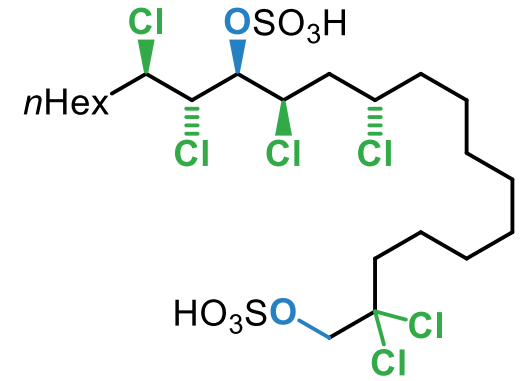


→ Toxizität Salzwassergarnelen: Cl/Br-Austausch kein Effekt
 Cl/F-Austausch Verminderung Toxizität

- Naturstoffklasse **Chlorsulfolipide**
- Synthesestrategien
- Beispiele Totalsynthesen

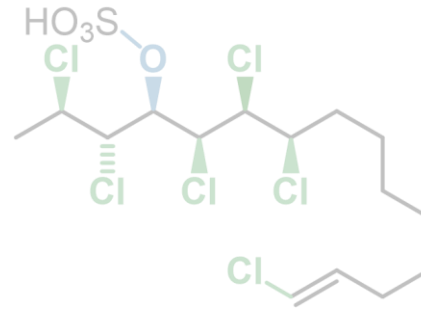


Mytilipin A

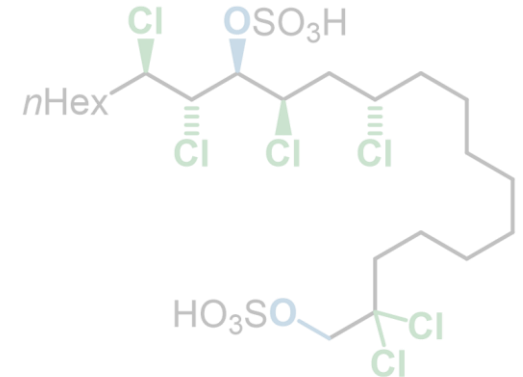


Danicalipin A

- Naturstoffklasse **Chlorsulfolipide**
- Synthesestrategien
- Beispiele Totalsynthesen

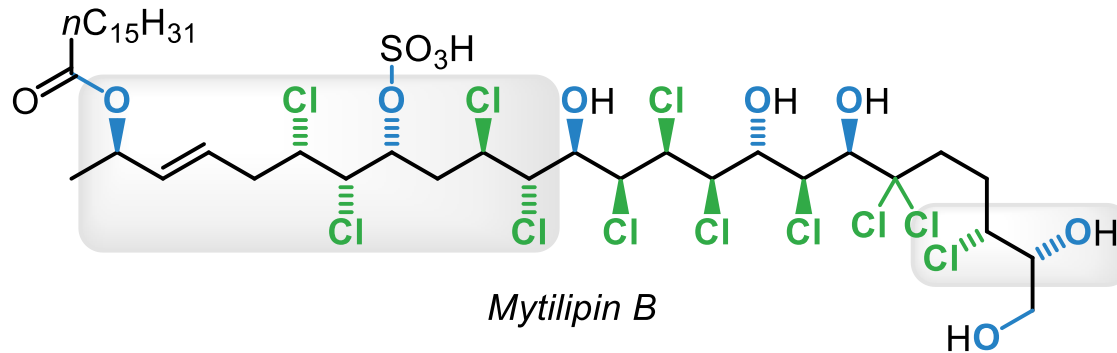


Mytilipin A



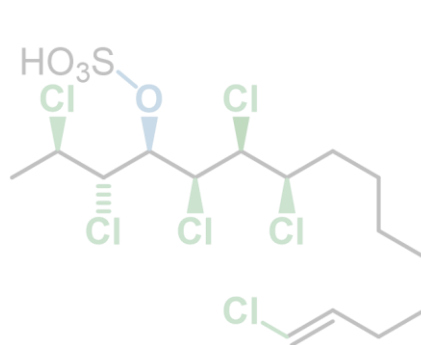
Danicalipin A

- **Strukturrevision durch Totalsynthese**

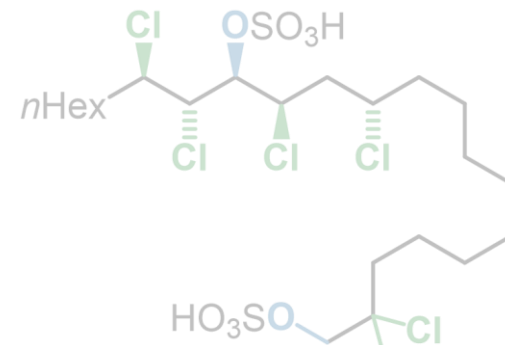


Mytilipin B

- Naturstoffklasse **Chlorsulfolipide**
- Synthesestrategien
- Beispiele Totalsynthesen

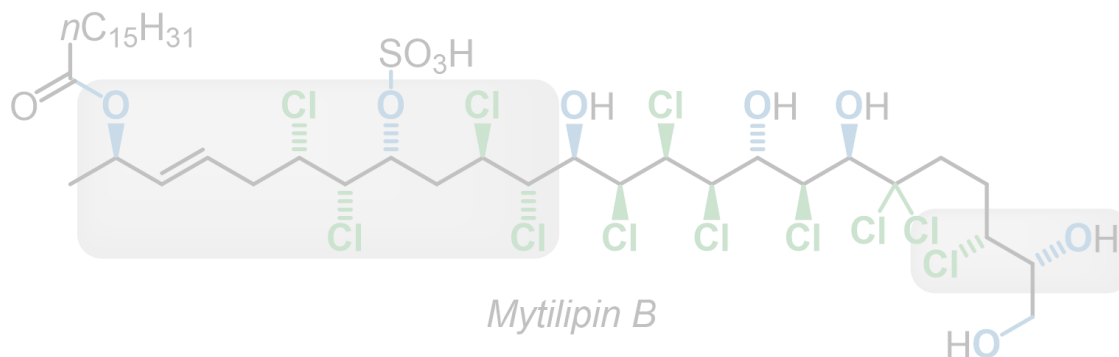


Mytilipin A

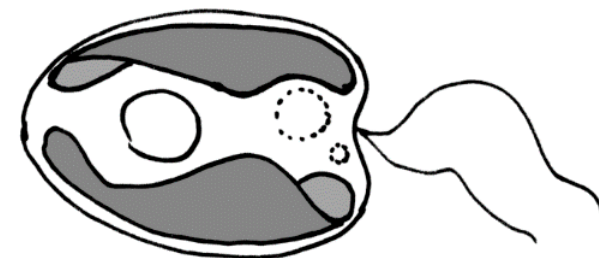


Danicalipin A

- Strukturrevision durch Totalsynthese



Mytilipin B



- Rigide Konformation → Bestimmung relative Konfiguration per NMR
- Struktur-Wirkungs-Beziehung:
Sulfolipide erhöhen Flexibilität der Zellmembran von einzelligen Mikroalgen
→ Essentiell für Fortbewegung

Former Coworker

- Tanja Stach (PhD)
- Ben Zoller (MSc)
- Pamela Hasanovic (BSc)
- Philipp Grewelinger (BSc)
- Marc Scheider (MSc)
- Christelle Mbouhom (BSc)
- Isabel Filbrich (MSc)
- Julia Dräger (BSc)
- Sebastian Motsch (MSc)



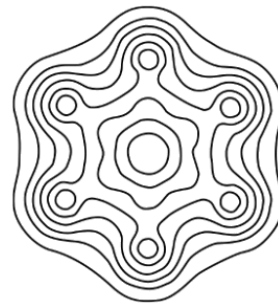
Mentors

- U. Kazmaier
- J. Jauch
- H.-G. Schmalz
- A. Koskinen
- B. List

Cooperation Partners

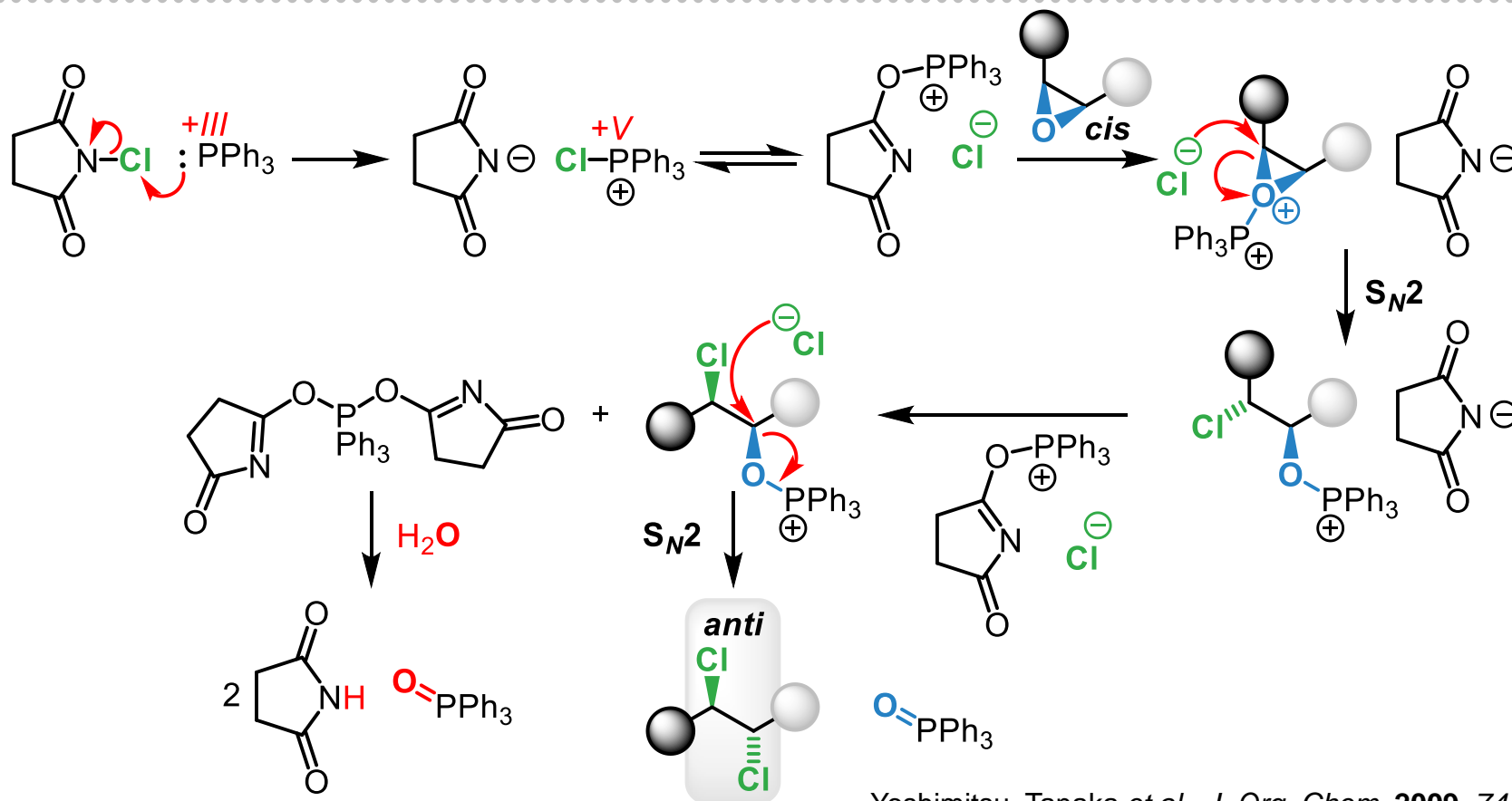
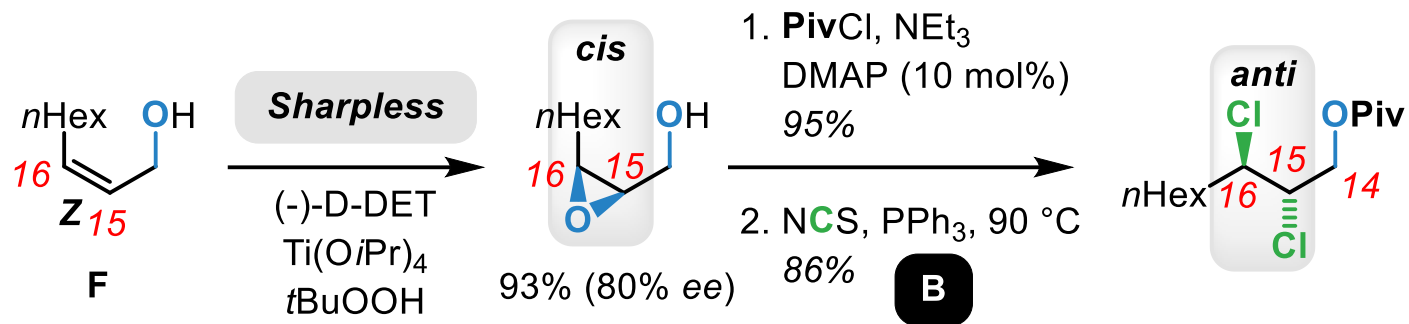
- A. Schäfer
- G. Hilt
- S. Heiles

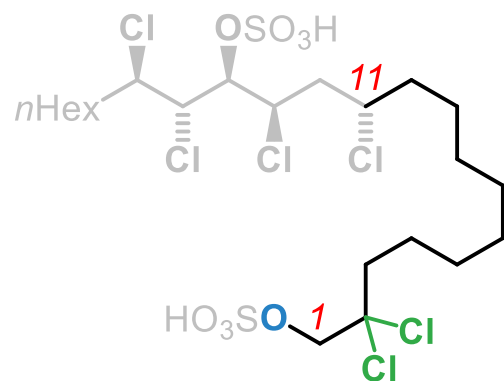
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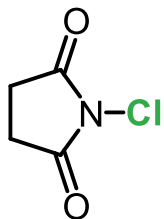
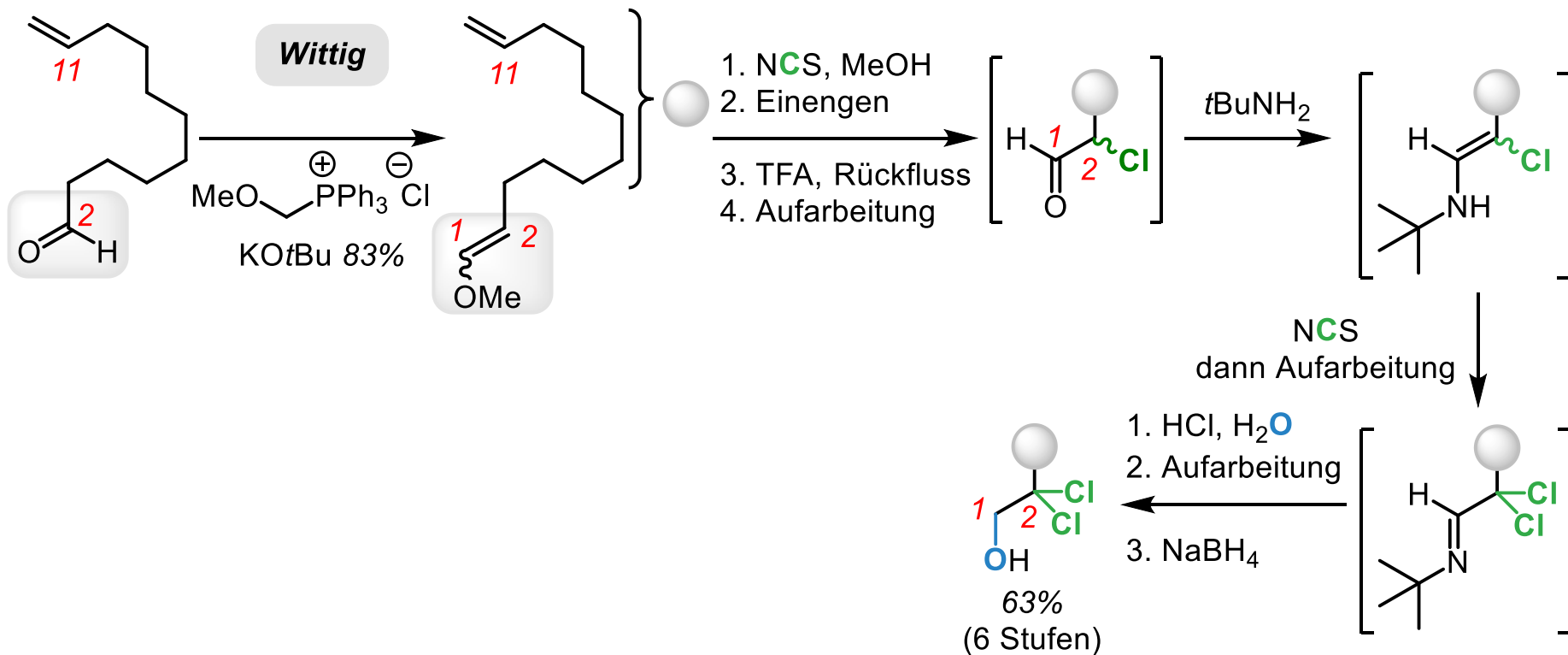


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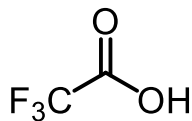




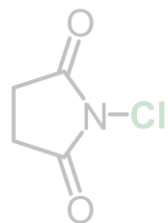
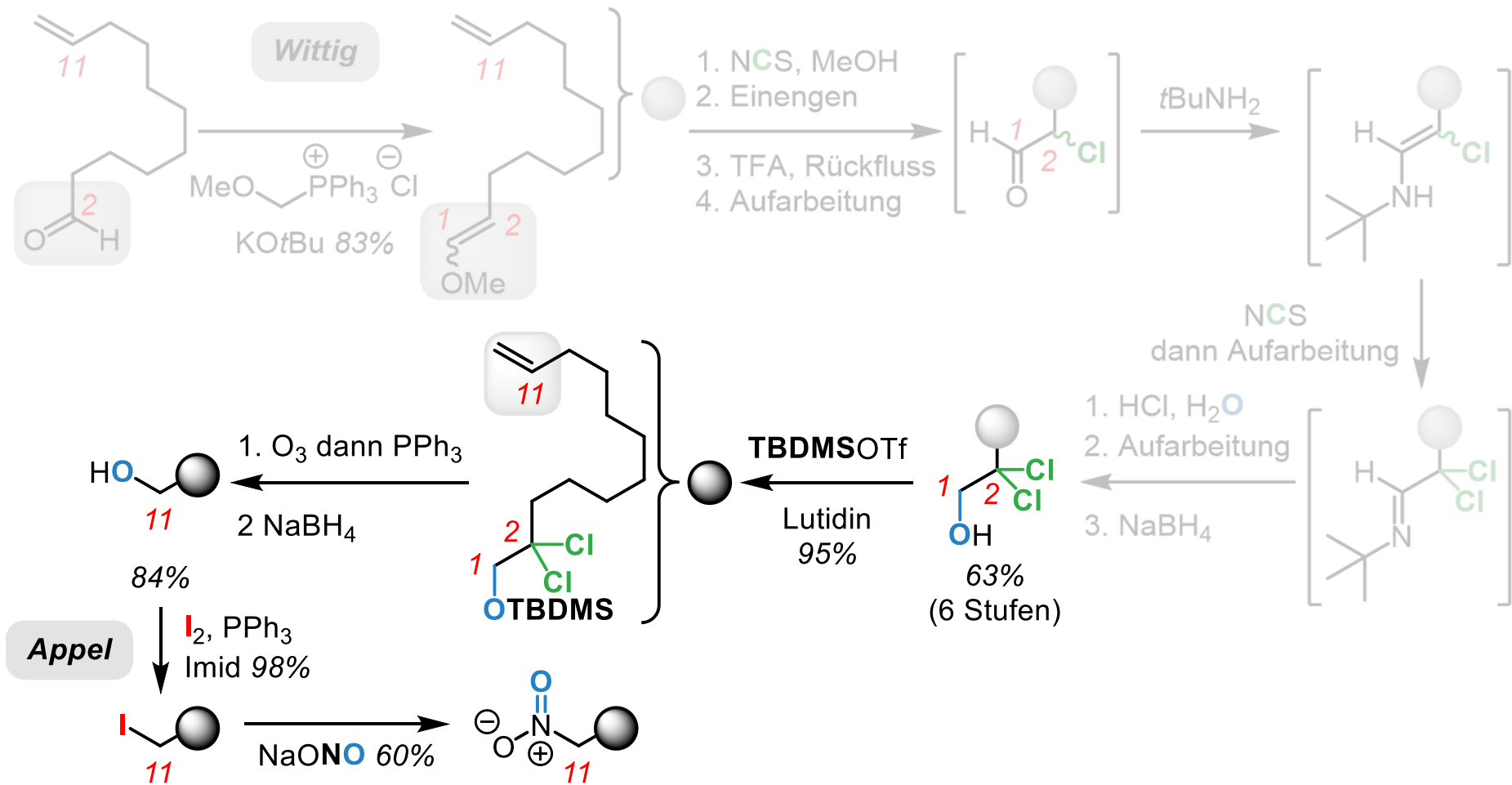




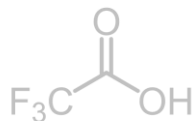
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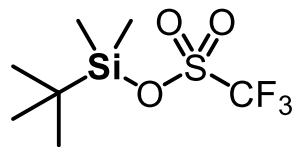
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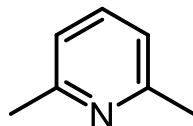
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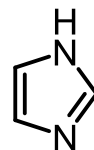
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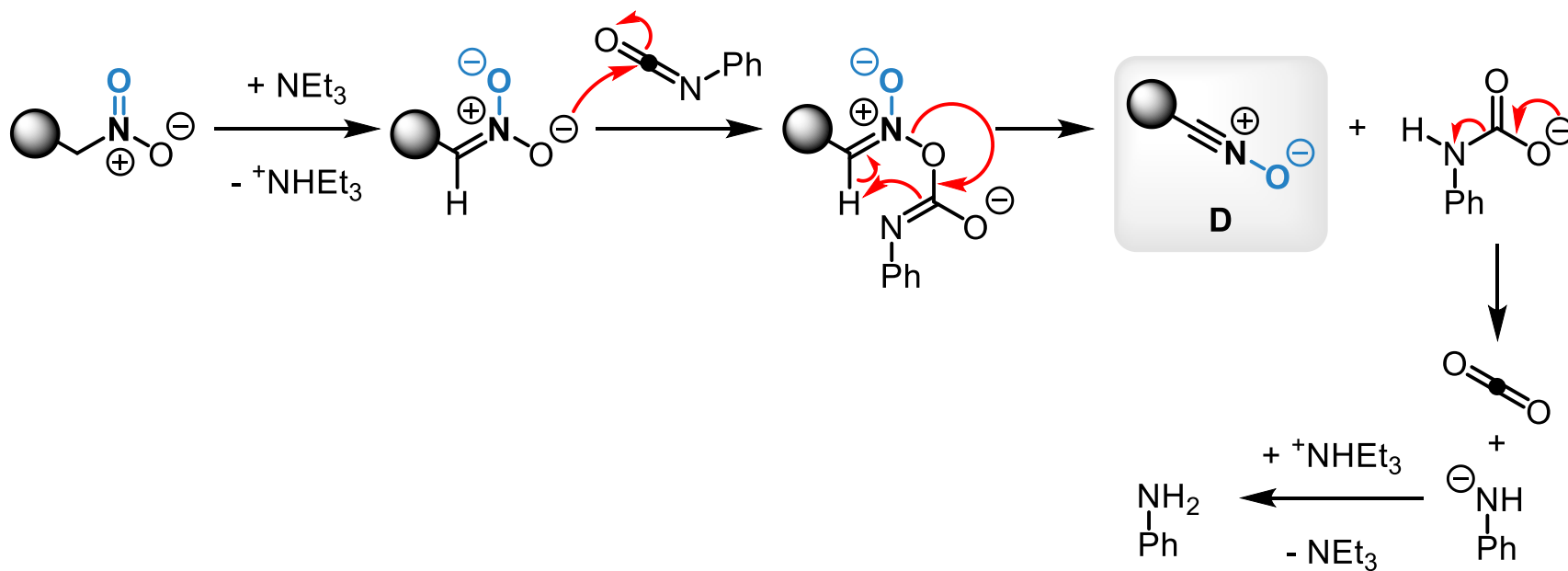
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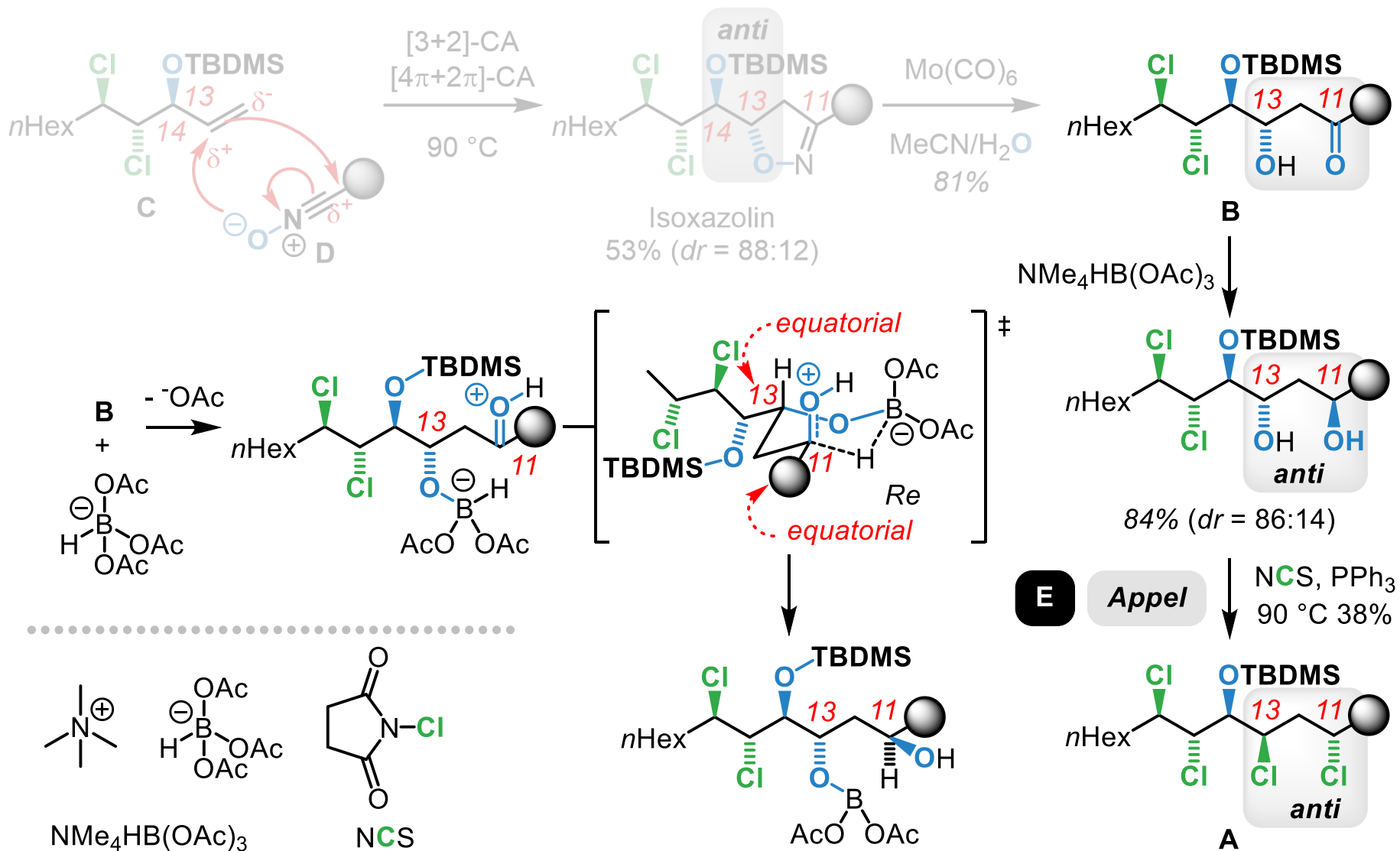


Lutidin



Imid





(a) Houk et al., *J. Am. Chem. Soc.* **1986**, 108, 2755. (b) Kobayashi et al., *Chem. Soc. Perkin Trans. I* **1985**, 1401.
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